

Habilitation Thesis

Polymers as Carriers of Active Agents

Polymery jako nosiče aktivních látek

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Field of Research: Technology of Macromolecular Substances

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SUMMARY

The habilitation thesis presents the topics of active polymer systems that could prevent the risk of microbial infections and contaminations. The introductory part reviews the potential forms of carriers, active molecules, as well as the characterization techniques of prepared systems. The second part is devoted to author's contribution to the subject covering the problems of antimicrobial polymer carriers based on both synthetic and natural polymers in the form of solutions, films, thin layers and nanofibrous membranes. The main goal of the commented research papers was to overview the approaches leading to the design of functional, cost effective and environmentally friendly polymer systems applicable in the food, cosmetic, medical or water treatment industry.

RESUMÉ

Habilitační práce se zabývá problematikou aktivních polymerních systémů, které mají sloužit jako prevence vzniku a šíření mikrobiálních infekcí. V úvodní části jsou představeny potenciální formy nosičů, typy aktivních látek a metody charakterizace připravených systémů. Druhá část je věnována diskuzi výsledků studií antimikrobiálních systémů na bázi syntetických i přírodních polymerů ve formě roztoků, filmů, tenkých vrstev nebo nanovlákenných membrán. Hlavní podstatou diskutovaných publikačních výstupů autora bylo zhodnotit přístupy vedoucí k vývoji funkčního, ekonomicky i environmentálně příznivého polymerního systému využitelného v potravinářství, kosmetice, zdravotnictví nebo procesech úpravy vody.

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INTRODUCTION

Microbial infections represent a serious problem, due to the persistence of pathogenic microorganisms present in various environments and applications, including especially pharmaceutical, food and cosmetic industry. Moreover, a number of bacterial strains become highly resistant to various kinds of drugs and are capable of facile spreading into different environments, such as air, soil and water. This is the reason why many of scientific groups constantly search for new possibilities and strategies in order to eliminate above-mentioned risks and thus prevent associated undesirable economic impacts. Some of up-to-know approaches suffer from various negative aspects, such as insufficient and shortterm efficiency, low stability, residual toxicity or high volatility. Application of active polymer materials is the perspective strategy how to fight with multi-drug resistant microorganisms and eliminate the above-mentioned drawbacks. These systems can be classified according to their mechanism of action to either passive of active type, which can be affected by many factors, such as molecular weight or polymer chain length. An emphasis is put on the character of base materials applied for the preparation of such systems, especially on their biodegradability, biocompatibility, non-toxicity and safety.

Presented habilitation thesis includes the results of author's scientific papers regarding the long-term goal consisting in the preparation and study of an effective and economically favourable antimicrobial material with potential applications in pharmaceutical, cosmetic, or food industry as active packaging, dressings, or for water treatment processes. The purpose of this work was to utilize the simple preparation methods for the design of the functional systems that are safe and do not promote bacterial resistance yet. The thesis also gives an overview of the polymer materials utilized as the carrier matrices, the potential active substances, as well as methods of preparation and characterization of the final systems, along with some critical issues associated with this topic. The carriers based on synthetic and natural polymers, namely poly (vinyl chloride), poly (vinyl alcohol), polyethylene oxide, poly lactide, and chitosan were of particular interest. The mutual interactions between the polymer matrix, active molecule and eventual "third" component (surfactant) affecting the stability were investigated.

1. LIST OF PUBLICATIONS

- P1 GALYA, T., SEDLARIK, V., KURITKA, I., NOVOTNY, R., SEDLARIKOVA, J., SAHA, P. Antibacterial poly(vinyl alcohol) film containing silver nanoparticles: preparation and characterization. *Journal of Applied Polymer Science* 110, 2008, 3178.
- P2 GALYA, T., SEDLARIK, V., KURITKA, I., SEDLARIKOVA, J., SAHA, P. Characterization of antibacterial polymeric films based on poly(vinyl alcohol) and zinc nitrate for biomedical applications. *International journal of Polymer Analysis and Characterization* 13, 2008, 24.
- P3 SEDLARIK, V., GALYA, T., SEDLARIKOVA, J., VALASEK, P., SAHA, P. The effect of hydrolysis degree on the properties of antibacterial polymeric films based on poly(vinyl alcohol) and zinc sulphate for biomedical applications. *Journal of Biomaterials Science* 21, 2010, 1421.
- P4 MERCHAN, M., SEDLARIKOVA, J., SEDLARIK, V., MACHOVSKY, M., SVOBODOVA, J., SAHA, P. Antibacterial polyvinyl chloride/antibiotic films: The effect of solvent on morphology, antibacterial activity and release kinetics. *Journal of Applied Polymer Science* 118, 2010, 2369.
- P5 MERCHAN, M., SEDLARIKOVA, J., VESEL, A., SEDLARIK, V., PASTOREK, M., SAHA, P. Characterization of antibacterial, mechanical, and structural properties of polyvinyl chloride/silver nitrate composites prepared by thermoplastic compounding. *International Journal of Polymer Analysis and Characterization* 15, 2010, 360.
- P6 MERCHAN, M., SEDLARIKOVA, J., FRIEDRICH, M., SEDLARIK, V., SAHA, P. Thermoplastic modification of medical grade polyvinyl chloride with various antibiotics: effect of antibiotic chemical structure on mechanical, antibacterial properties, and release activity. *Polymer Bulletin* 67, 2011, 997.
- P7 MERCHAN, M., SEDLARIKOVA, J., VESEL, A., MACHOVSKY, M., SEDLARIK, V., SAHA, P. Antimicrobial silver nitrate-doped polyvinyl chloride cast films: Influence of solvent on morphology and mechanical properties. *International Journal of Polymeric Materials* 62, 2013, 101.
- P8 SEDLARIKOVA, J., DOLEZALOVA, M., EGNER, P., PAVLACKOVA, J., KREJCI, J., RUDOLF, O., PEER, P. Effect of oregano and marjoram essential oils on the physical and antimicrobial properties of chitosan based systems. *International Journal of Polymer Science* 2017.
- P9 KOLAROVA RASKOVA, Z., STAHEL, P., SEDLARIKOVA, J., MUSILOVA, L., STUPAVSKA, M., LEHOCKY, M. The effect of plasma pretreatment and cross-linking degree on the physical and antimicrobial properties of nisin-coated PVA films. *Materials* 2018, 11(8), 1451.

- P10 SEDLARIKOVA, J., JANALIKOVA, M., RUDOLF, O., PAVLACKOVA, J., EGNER, P., PEER, P., VARADOVA, V., KREJCI, J. Chitosan/thyme oil systems as affected by stabilizing agent: physical and antimicrobial properties. *Coatings* 2019, 9, 165.
- P11 KOUSAL, J., KRTOUS, Z., KOLAROVA-RASKOVA, Z., SEDLARIKOVA, J., SCHAFER, J., KUCEROVA, L., SHELEMIN, A., SOLAR, P., HURAJOVA, A., BIOEDERMAN, H., LEHOCKY, M. Degradable plasma polymer films with tailored hydrolysis behavior. *Vacuum* 2019, 173, 109032.
- P12 KOUSAL, J., SEDLARIKOVA, J., KOLAROVA-RASKOVA, Z., KRTOUS, Z., KUCEROVA, L., HURAJOVA, A., VAIDULYCH, M., HANUS, J., LEHOCKY, M. Degradable poly(ethylene oxide)-like plasma polymer films used for the controlled release of nisin. *Polymers* 2020, 12, 1263.
- P13 PEER, P., SEDLARIKOVA, J., JANALIKOVA, M., KUCEROVA, L., PLEVA, P. Novel poly vinylbutyral/monoacylglycerol nanofibrous membrane with antifouling activity. *Materials* 2020, 13, 3662.

2. THEORETICAL BACKGROUND

2.1 Types of carriers

In the area of polymer active systems, types of carriers play an important role for subsequent applicability. Carriers for active molecules can be prepared from many types of materials and can exist in different forms, such as micro/nanoparticles, emulsion systems, polymer films, fibers, micelles, liposomes etc. A number of factors have to be taken into consideration, especially the character of active substance, potential mutual interactions with the carrier, and/or required properties and application. Optimum supporting system should be stable, safe, and economically, as well as ecologically favourable.

2.1.1 Micelles

Micelles are colloid self-assembled aggregates formed by amphiphilic surface active compounds, called surfactants, above the critical micelle concentration (CMC). Surfactant molecules contain two parts, hydrophilic head and hydrophobic tail. Micellar aggregates can exist in different forms, some types are shown in Fig. 1 [1].

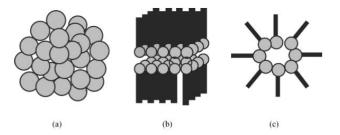


Fig. 1. Types of aggregates: a) spherical, b) lamellar, c) reversed micelle [2]

The standard Gibbs free energy of non-ionic micelle formation per mole of surfactant is given by the Equation 1.1:

$$\Delta G_m^0 = RT \ln(CMC) \tag{1.1}$$

Solubilization capacity of surfactant micelles is determined by the molecular structure of amphiphile, as well as solubilized molecule, mechanism of entrapment and physicochemical interactions of present constituents [1]. In aqueous solutions, hydrophobic agents, e.g. drugs, are incorporated into hydrophobic core of micelles, which enhances their solubility. On the other hand, polar materials are commonly located in so-called palisade layer of the micelle (Fig. 2) [2].

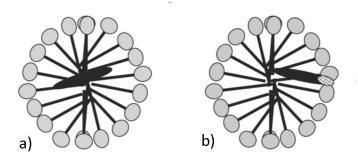


Fig. 2 Solubilization in micelle a) hydrophobic substance in core, b) hydrophilic substance in palisade layer [2]

Through the solubilization into micelles, quite large quantities of poorly soluble active agents can be incorporated in polar solution. Moreover, due to low water concentration in micellar core, hydrolytic degradation is suppressed, which increases the chemical stability of solubilized molecule. Release kinetics can be controlled, thanks to partitioning of encapsulated agent toward the micelle, and the effect of active molecule in desired target is enhanced [3].

Block copolymer micelles

A special type of aggregates is represented by polymer micelles based on self-assembly of block copolymers above the critical micelle concentration. These diblock or triblock copolymers may form not only micelles in dilute solutions, but also a number of liquid crystalline phases. In polar media, these nanostructures contain external hydrophilic corona and hydrophobic core, whereas in an opposite arrangement exists in nonpolar solvents (Fig. 3).

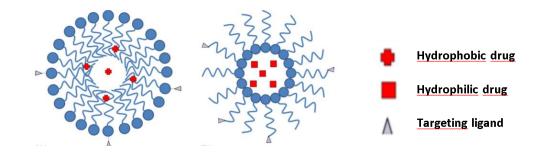


Fig. 3 Structure of polymer micelle in a) polar and b) nonpolar media [4]

Particularly, poly(ethylene oxide)/poly(propylene oxide) copolymers (PEO/PPO) have been intensively studied during the last decade for potential applications as drug delivery systems, especially for their commercial accessibility and variability of molecular weights. Similarly to low molecular weight non-ionic surface active agents, micelle formation of block copolymers is

supported by an increasing length of hydrophobic part (PPO) and decreasing proportion of hydrophilic part (PEO) of molecule.

Micellization of PEO/PPO block copolymers is significantly affected by temperature. Because of lower solubility of PEO blocks at higher temperature, the process of micelle formation is promoted. As in case of common low molecular weight aggregates, the location of solubilizate is primarily affected by its properties. More hydrophobic molecule will tend to organize into the micelle core, whereas more amphiphilic agents will preferably locate in the palisade layer.

Triblock copolymers of A-B-A type, known under the trade name Pluronic or Lutrol, represent an example of polymer surfactant suitable for drug delivery system. Such structure has hydrophobic PEO based core ended with hydrophilic blocks of PEO. In the work of Chaudhari et al. [5], a solubilization of poorly water soluble drug Lamotrigine into Pluronic based aggregates was investigated. A high entrapment efficiency was achieved as well slower release rate of a model drug from Pluronic mixed micelles. The micellization process of Pluronic copolymers as the potential carriers of active agents has been investigated during the last two years in the author's workplace within the proceeding diploma works. Three types of Pluronics of various molecular weights were selected to study their micellar characteristics with the presence or absence of phenolic compounds, such as curcumin, essential oils or their components [6].

2.1.2 Emulsion/dispersion based systems

Emulsions are dispersion systems composed from two immiscible liquids, of which one represent the internal or discontinuous phase that is distributed in the second continuous phase. Emulsions are widely applied in food industry, pharmacy and cosmetics. They can be classified according to the polarity to oil in water (O/W) and water in oil (W/O) emulsions, however, even multiple emulsion types (e.g. O/W/O or W/O/W) exist. Based on the size of droplets, there are macroemulsions (1 to 100 μ m), nanoemulsions (20 to 200 nm) and microemulsions (10 to 100 nm). Under homogenization procedure, the concentration of an emulsifier, as well as a co-surfactant play an important role here, too [7].

Microemulsions and nanoemulsions

Microemulsions are transparent, thermodynamic stable isotropic dispersion systems containing the particles ranged from 10 to 100 nm. They are formed spontaneously with a higher portion of co-surfactant that causes further decrease of an interphase tension. Nanoemulsions usually contain particles in the size from

20 to 200 nm with high interfacial area and exhibit a high solubilization capacity and low viscosity. Compared to microemulsions, they are not thermodynamically stable but they can possess a high portion of kinetic stability against sedimentation, flocculation and/or coalescence when prepared under optimum conditions [8]. The formation of nanoemulsions can occur by high or low energy homogenization methods. The latter mentioned group includes techniques utilizing the phase inversion during emulsification, such as emulsion inversion point (EIP) or phase inversion temperature (PIT) method [7].

Surfactants represent the important components in emulsion systems. They can be classified according to the hydrophilic part to non-ionic and ionic types, which are further divided into three subgroups to anionic, cationic and amphoteric. Anionics are most often represented by carboxylates (R-COO-M+), sulphates (R-OSO₃-M+) and sulphonates (R-SO₃-M+). Due to the dissociation degree, quaternary ammonium compounds (R₄N+X-) are the most frequently applied types of cationic surface active agents. Amphoteric surfactants containing both cationic and anionic part in the hydrophilic head are represented by betaines. Non-ionics are not charged but the solubility in polar media is ensured by the presence of functional groups, such as polyoxyethylene (R-OCH₂CH₂O-) [2].

Surfactants play important roles in the emulsion formation, as well as their stability. They decrease the interfacial tension γ , which leads to the reduction of droplet size in emulsions. In practical applications, the mixtures of surfactants are commonly used, because of their synergetic effects leading to a more significant decrease of the interfacial tension γ . It is known that different components provide varying surface activity, and the compounds with the lowest γ are preferentially located at the interface. Even the mixtures of polymers and surfactants exhibit some synergetic effects in the surface activity [9]. Surfactants also support the existence of interfacial tension gradients, which allows the stabilization of the system. This phenomena is associated with so called Gibbs-Marangoni effect describing the processes occurring at the interface between emulsion droplets. The depletion of surface active agent in the thin film between approaching droplets leads to the gradient in the interfacial tension γ . As the consequence, an inward flow of liquid together with the surfactant molecules begins and thus, the droplets are driven apart and the system is more stable. The Bancroft rule stating that the continuous phase is formed by that, in which the surfactant is more soluble, can be explained by the Gibbs-Marangoni effect, too. Then, surfactants having hydrophilic-lipophilic balance (HLB) > 7, tend to form O/W emulsions, while surfactants with HLB < 7 prone to form W/O emulsions [2].

Lipid particles

Lipid nanoparticles are nanoemulsion based systems of O/W type, in which a liquid oil is replaced by a solid lipid (solid lipid nanoparticles, SLN) or a mixture of solid and liquid lipids (nanostructured lipid carriers, NLC). The average size of SLN is in the range from 40 to 1000 nm and the surfactant concentration from 0.5 to 5 wt% is used for the stabilization. These carriers can be produced by different techniques, such as high-pressure homogenization, microemulsion technique, solvent injection (or displacement) method, phase inversion or ultra-sonication.

SLN have the advantages in the entrapment of both hydrophilic and hydrophobic active agents at high concentrations. However, a drug loading capacity is lower because of the lipid crystalline structure. NLC possess a less ordered structure thanks to the presence of liquid and solid lipid mixture [10].

2.1.3 Liposomes

Liposomes are nowadays widely applied in pharmaceutical, cosmetic and food industry. These spherical vesicles have hydrophilic core surrounded by at least one phospholipid-based bilayer. The most commonly applied phospholipid is phosphatidylcholine. Liposomes can be classified according to the size and structural type to:

- small unilamellar vesicles (SUV): 20 100 nm;
- large unilamellar vesicles (LUV): > 100 nm;
- giant unilamellar vesicles (GUV): > 1000 nm;
- oligolamellar vesicles (OLV): 100 500 nm;
- multilamellar vesicles (MLV): > 500 nm.

Liposomes can entrap hydrophilic, hydrophobic and even amphiphilic molecules, despite the fact that their core itself is hydrophilic. While hydrophobic agents are incorporated between the lipid bilayer, hydrophilic molecules are encapsulated in the core. Amphiphilic substances remain at the water/lipid interphase based on their affinity to the liposome parts [10, 11].

Liposomes represent perspective candidates for delivery systems due to their biodegradability, biocompatibility, low toxicity and easy preparation methods. A rapid release of poorly water-soluble drugs encapsulated in the lipid bilayer is one of the negative aspects. Another drawback consists in low physical and chemical stability caused by the oxidation and hydrolysis of phospholipids [10]. Liposomes can be prepared by the dispersion of lipid molecules in an aqueous phase carried out by mechanical methods, such as film hydration method, sonication,

microfluidization, extrusion, or by the replacement of organic solvents, such as proliposome-liposome method, ethanol injection, and lyophilisation [12].

2.1.4 Niosomes, transfersomes and ethozomes

Niosomes are known from the seventies when they were developed for the utilization in cosmetic industry. At present, also the applications in the pharmacy exist. Niosomes are biodegradable and biocompatible vesicles based primarily on non-ionic surfactants and might contain cholesterol or its derivatives, too. Cholesterol serves as an additive substance able to interact with hydrophilic part of non-ionic surface active agent. Some of niosome properties, such as entrapment efficiency, stability (storage time) and release conditions, can be affected by its incorporation [13]. Stability of niosomes might be also enhanced by the addition of charged molecules that prevent aggregation mechanisms and improve skin permeation properties. Non-ionic molecules based vesicles are resistant to oxidation and high temperatures. Moreover, niosomes are less expensive when compared to liposomes. Niosomes might be easily prepared by thin layer hydration technique, the organic solvent injection method or e.g. the reverse-phase evaporation method [10].

Transfersomes were firstly developed at the beginning of nineties. These deformable vesicles contain phospholipids and an edge activator, which is often represented by a single chain surfactant with a high curvature radius that is able to lower the stiffness of a bilayer. Consequently, the vesicles gain an elastic structure resistant to the rupture. While phospholipid is the primary constituent, surfactant occupies from 10 to 25 % in the structure, and the solvent is present in the concentration from 3 to 10 %. Transfersomes can accommodate various types of molecules, such as peptides or antioxidants. They may be prepared by simple methods, including the combinations of vortexing-sonication and rotary evaporation-sonication [10].

Ethosomes are phospholipid-based vesicles with high ethanol content (20–25 %) that were developed in 1997. Similarly to transfersomes, these vesicles are significantly deformable with high ability to permeate into the skin. Included ethanol serves as an agent for enhancing the permeation, due to its ability to affect the bilayer structure of *stratum corneum*. Ethosomal vesicles enable to deliver the encapsulated molecule deeper in the skin in comparison to liposomes. Additionally, ethosomes are smaller with better entrapment efficiency and stability. On the other hand, they are instable because of the oxidative degradation. Ethosomes can be prepared by the convenient hot and cold methods, or e.g. by classic mechanical dispersion method [14]. The scheme of discussed structures is shown in Fig. 4.

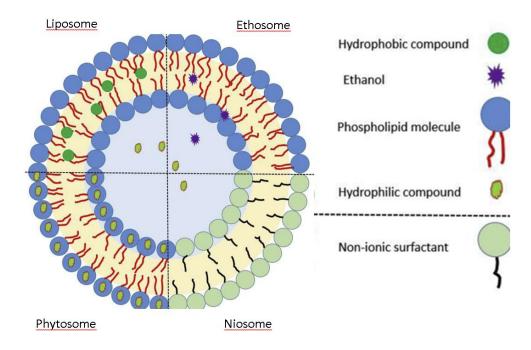


Fig. 4 Scheme of selected vesicle-based systems [15]

2.2 Polymer systems

Polymer materials have found applications in a wide range of disciplines including pharmaceutical, agricultural, food and cosmetic industry. Polymers can be classified according to the structure to linear, branched and cross-linked types. Based on their origin, they are divided into natural and synthetic materials. The latter mentioned group comprises of the polymers prepared in the laboratory, such as polyethylene, polystyrene, polyvinyl chloride, synthetic rubber etc. Natural polymers are isolated from natural materials (plant and/or animal sources) and are of great interest nowadays due to the general trend of eliminating the amount of synthetic substances in environment. The representatives include polysaccharides, proteins, nucleic acids and natural rubber.

2.2.1 Antimicrobial polymers – mechanisms of action

Active polymer materials have been under extensive research in order to reduce the risk of microbial contamination that represent serious problem especially in healthcare products, food applications or water treatment systems [16]. Hence, several methods have been developed to obtain polymers with desired antimicrobial properties. Polymer systems with bioactive compounds can be found in the form of coatings, films, micro- or nanospheres, nanogels, or e.g. capsules [17]. Considering the function of antimicrobial polymers, they can work either by passive or active way. Passive action includes a material passively preventing adhesion of bacteria, not actively interacting with them. Passive polymers are mostly hydrophilic, having low surface free energy, or negatively

charged. Polyphenol or poly(ethylene glycol) based polymers have been extensively studied as passive polymers preventing the growth of Gram negative and Gram positive bacteria [19,20]. Active polymers are based on the active action when bacteria adhering to the polymer surface are killed. In this case, functional active agents play a predominant role. The examples of active polymers are poly(iminoethylene), poly(guanidine) and positively charged quaternary ammonium types [22].

Antimicrobial polymers can be classified into polymer biocides, biocidal polymers and biocide-releasing polymers consisting of polymers enriched with active molecules (Fig. 5).

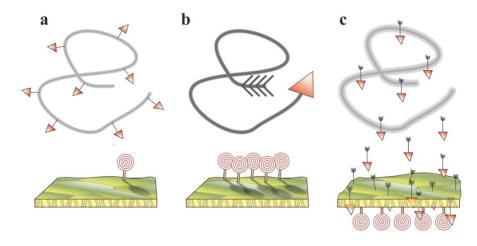


Fig. 5 Types of antimicrobial polymers: a) polymer biocides, b) biocidal polymers, c) biocide-releasing polymers [21]

Polymer biocides include materials consisting of repeating bioactive molecules, such as hydroxyl, carboxyl or amino groups that are covalently linked. However, the process of polymerization of biocidal monomer units does not always lead to the production of active polymer, caused either by an insufficient solubility in water or because the active substances are not able to reach the target site. The examples of polymer biocides include polymers with quaternary ammonium and phosphonium salts or benzimidazole derivatives [22].

In case of biocidal polymers, the active site is represented by the macromolecule itself and no repeating bioactive unites are required. Since many microbial organisms carry a negative charge, polycations are attracted to their surface, which results in the disruption of their outer membrane leading to the cell lysis and death. Therefore, many types of biocidal polymers contain cationic groups, such as quaternary ammonium, phosphonium, tertiary sulfonium, or guanidinium. Chitosan and its derivatives have been studied, mainly due to their advantageous properties including biodegradability, biocompatibility and absence of toxicity. Chitosan's antimicrobial properties are strongly governed by its pH value. Generally, when pH value is less than pKa, amino groups are in protonated

form and can interact with the cell wall. On the other hand, when pH is above pKa, chitosan can operate as antimicrobial due to hydrophobic interactions.

The third group comprises so called biocide-releasing polymers that can proceed through either polymerization of biocide-releasing molecules to polymeric backbone, or the polymer/biocide-releasing molecules composites. The polymer material serves as a carrier system for active molecules, such as antibiotic or antiseptic compounds. Systems for controlled release have been intensively studied during last years as the innovative principle in reducing the risk of microbial infections by the targeted delivery. A great effort has been devoted to study of the release kinetics and influential factors [22].

2.2.2 Modification of polymers with antimicrobial substances

Antimicrobial polymers can be prepared by several methods, including deposition of active agent on the substrate surface, such as vapour, sputter or ion beam coating, or plasma treatment. The last mentioned technique was applied in the author's publications **P9**, **P11** and **P12**. Also, dip or spin coating can be utilized as an alternative approach to minimize bacterial contamination. However, coating or immersion of the material in solution of an antibacterial substance suffers from many limitations, including rapid release of active substance in a short time. Another possibility includes an incorporation of antimicrobial substance into polymer matrix, e.g. be melt blending or solvent casting technique. Melt compounding has been investigated in **P5** and **P6** using polyvinyl chloride as the base matrix and organic or inorganic active molecules to get the systems with antibacterial activity. The latter method, solvent casting, has been utilized in most of the author's publications (**P1–P4**, **P7**, **P8**, **P10**) investigating both synthetic and natural polymers as potential carriers of active agents.

2.2.3 Antifouling surfaces

Antifouling surfaces are used to eliminate the consequences of biofouling mechanisms occurring through the colonization by microorganisms resulting in the formation of biofilms on the exposed surface. To enhance the antifouling properties, the surface modification, either by coating a thin film, or by grafting of polymer chains on the surface can be carried out (Fig. 6).

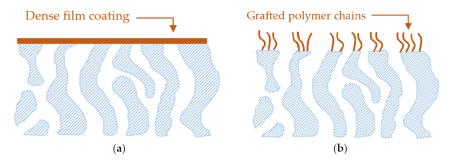


Fig. 6 Modification of polymer surface by: a) thin film coating, b) grafting of polymer chains [23]

It is necessary to realize that materials selected for these processes (coating or grafting) are not affine to potential foulants, which leads to the limitation of any undesirable interactions between them and the substrate. The antifouling properties are also affected by the surface roughness, wettability and charge. Rough surfaces are more prone to spontaneous fouling because of the increased deposition of the foulants in the "pockets" of the surface. If the surface is charged, it can attract the foulants with counter ions due to the electrostatic interactions [23]. Therefore, the knowledge on the surface properties, such as wettability and contact angle (reported on in the chapter 2.2.4), is crucial to understand this topic in a broader context. Antifouling membranes based on poly(vinyl butyral) enriched with monocaprin were designed in the author's publication **P13**.

2.2.4 Characterization methods of active polymer systems

When active polymer systems, both film forming polymer solutions, and films or thin layers, are prepared, various techniques for their characterization might be used. A range of these is employed for evaluation of the short or long-term stability, so that the products fulfil the requirements of practical applications. Stability can be understood as the ability to resist the external factors leading to the structural changes of the product. It is known that all the systems tend to retain the most energetically advantageous state. The following chapters include the selection of methods used within the author's research for the characterization of active polymer systems.

Particle size

Particle size belongs among the important characteristics giving evidence of the physical stability of the solution, as well as affecting the texture and appearance of the resultant system [24]. A dynamic light scattering (DLS) technique can be utilized to measure the size of particles, which is based on the analysis of the intensity of light scattered from solution or suspension. DLS (also known as photon correlation spectroscopy – PCS) measures so called Brownian motion (random thermal motion of fine particles and molecules). Since smaller particles move faster than larger ones, there is a relationship between the particle size and its speed defined by the Stokes-Einstein equation (1.2) [25]:

$$D = \frac{kT}{6\pi\eta R_H} \tag{1.2}$$

Where D is the translational diffusion coefficient, η is the viscosity, k is Boltzmann's constant, T is the absolute temperature, R_H is the hydrodynamic particle radius.

Electrokinetic measurement

Zeta (ζ) potential of liquid dispersions informs on an electrostatic charge present on the slipping plane between fixed and diffuse region of the electric double layer. It depends on the physichochemical properties of systems and can affect the processes of dispergation, adsorption and/or aggregation. Generally, particles possessing a sufficiently positive (>+ 30 mV) or negative (<- 30 mV) zeta potential, repel each other, which results in an increase of stability. On the contrary, a significant decrease in stability is monitored when zeta potential value ranges from 0 to \pm 10 mV due to a higher potential to aggregation [26].

The electrokinetic measurement can be also carried out on the solid surfaces and is associated with the dissociation of surface groups, the adsorption of cations or anions, or polyelectrolytes, or electron depletion. This can be measured either by electrophoretic methods, or the streaming potential can be determined when a relative movement of the solid or liquid phases is performed to generate an electrical potential or current. The latter mentioned is often applied for the measurement on planar surfaces [27].

Viscosity measurement

Rheological properties play an important role, especially for the polymer solutions intended for the processing by electrospinning that is discussed in Chapter 3.3. A direct relationship between the rheological behaviour and the type and concentration of polymer was proved. Therefore, the viscosity is being studied within the most of papers investigating the electrospun nanofibers [28].

The viscosity of polymer solutions is given by the Huggins equation (Equation 1.3):

$$\eta = \eta_0 [1 + [\eta]c + k_H [\eta]^2 c^2 + \cdots]$$
 (1.3)

Where c is the concentration of the polymer solution with viscosity of η , η_0 is the solvent viscosity, $[\eta]$ it the intrinsic viscosity, and k_H is the Huggin's coefficient. There is the direct relation between the intrinsic viscosity and the size and shape of the molecule in solution, and thus, the molecular weight. The Huggin's coefficient is the function of the molecular interactions.

A following expression (Equation 1.4) indicating the reduced viscosity can be acquired after editing the above equation:

$$\eta_{red} = [\eta] + k_H [\eta]^2 c \tag{1.4}$$

When the reduced viscosity is plotted versus the concentration of the solution, a linear dependence with the intrinsic viscosity as the intercept is obtained.

The intrinsic viscosity can be used for the calculation of the molecular weight applying the Mark-Howink equation (1.5):

$$[\eta] = K M_V^a \tag{1.5}$$

Where K and a are the constants for the selected pair of polymer and solvent [29].

Surface tension measurement

When the surfactant is included in the system, the measurement of surface tension plays a significant role. Surface tension of liquids is the result of attractive forces between the molecules. While the molecules present in bulk liquid exhibit the same attractive forces in all directions, the surface molecules are in asymmetric force field, which leads to so called surface energy and/or surface tension (Fig. 7) [29].

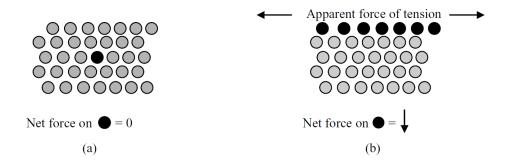


Fig. 7 Attractive forces acting on the molecule (a) in the bulk, (b) at the interface [2]

The surface tension can be also identified as a force acting at the surface related to the length, thus, it is expressed in N/m, or mN/m. It is known that surfactants significantly reduce the surface tension even at very low concentrations, until the critical micelle concentration (CMC) is reached, after which it is more or less constant (Fig. 8). This is because activity of monomers is constant and micelles are not surface active. Therefore, the break in surface tension relates more to the processes occurring in solution, rather than at the surface. The efficiency of a surfactant is determined to characterize its amount necessary to achieve a certain reduction in surface tension, usually set to 20 mN/m (the corresponding surfactant concentration is indicated as c_{20}). On the other hand, the effectiveness of the surfactant is identified as the reduction of surface tension at the critical micelle concentration (γ_{CMC}) [29].

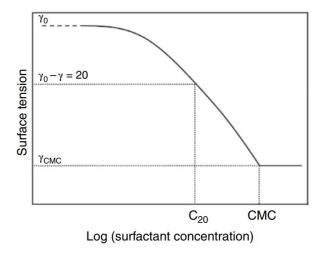


Fig. 8 Surface tension vs. the surfactant concentration logarithm [29]

The surface tension decrease is caused by the strong adsorption of surfactants at the surface. Gibbs defined the surface as the locality where the concentration of solvent is half between that in the solvent and the vapour. The relationship between the adsorption and surface tension is given by the Gibbs equation (Equation 1.6):

$$\Gamma = -\frac{1}{nRT} \frac{d\gamma}{d\ln a} \tag{1.6}$$

Where R is the gas constant, T is the absolute temperature, Γ is the surface excess (moles adsorbed per unit area of an interface), a is the surfactant activity in the bulk solution, and n is the constant. The concentration of surface active agent is usually very low below CMC value, therefore, the surfactant activity a can be replaced by its concentration c. Then, the surfactant adsorption can be determined from the slope of a dependence of the surface tension vs. concentration logarithm [27].

The surfactant adsorption is being studied at the author's workplace within the qualification student works, mostly focused on block copolymer non-ionic surfactants, or their mixtures with less common types of ionic surface active agents, such as sodium dioctyl sulfosuccinate (DSS), that has one sulphonate head group and two hydrophobic chains, and is able to form reversed micelles.

The surface tension can be measured by different techniques including capillary elevation, stalagmometry, or Wilhelmy plate method, which was applied in the auhor's publications (**P8**, **P10**, **P13**). Within this technique, a thin plate with defined dimensions is vertically immersed into the liquid and the acting force, which is related to the surface/interfacial tension and contact angle, is measured (Equation 1.7).

$$\gamma = \frac{F}{L \cdot \cos \theta} \tag{1.7}$$

Where L is the wetted length of plate, equal to its perimeter (thickness is small that can be neglected), F is the maximum force achieved just before the plate detachment from the liquid surface.

Microscopic analysis

Surface and interface characterization can be performed by several microscopic techniques, such as light microscopy in reflection or transmission, interference microscopy for surface topography, brewser angle microscopy, surface plasmon microscopy, fluorescence microscopy etc.

With the optical microscope, the state of polymer dispersion can be evaluated, i.e. if the particles are uniformly dispersed, coagulated or weakly flocculated. Brownian motion can limit the accuracy of analysis, thus, the samples have to be dried before the measurement, if it is possible. After the obtaining the image, it is analysed either by hand, or, the most often at present, by the appropriate computerized image-analysis system. In case of spherical particles, the particle diameter can be simply determined. However, the situation is more complicated for non-spherical particles, when various dimensions have to be defined and a calibration is required [27].

Transmission electron microscopy (TEM) can be used for the imaging and size analysis at significantly high resolution. A dried sample placed on the grid is then inserted in the microscope column that is evacuated. Afterwards, the beam of electrons is applied to produce an image that is focused onto a fluorescent screen. The wavelength of electron bean is driven by the accelerating voltage, with 10 or 20 kV. A stronger scattering occurs for the particles of high-atomic-number elements. Particles ranged from 10 nm to 10 µm can be analysed by this technique reliably. Scanning electron microscopy (SEM) is applied to obtain the surface image at nanometre lateral resolution due to electron absorption or emission. A sample is mounted on an aluminium stub usually coated with a thin conductive material that is sputtered at a specific angle, in order to prevent the electrostatic charge to interfere with the image. The resolution is lower than in case of TEM and is limited by the size of the scanned spot [27].

Mechanical properties

Mechanical properties, such as tensile strength, elongation at break, puncture strength, or puncture deformation, belong among the important characteristics of polymer films. Analysis of the mechanical properties can be carried out by

different methods, including common tensile test measuring of Young's modulus, or e.g. bulge test for determination of biaxial modulus and residual stresses. To measure the polymer film deformation, the techniques like the laser interferometry, atomic force microscopy, or mechanical profilometry can be used. Since all these techniques are very sensitive to vibration and noises, the resolution can be limited [30].

For measuring mechanical characteristics, various types of equipment can be applied, depending on the type and size of the sample. Texture analysis machine can be utilized even for the measurement of a large range of parameters providing the complex testing applications. For example, in case of edible polymer films intended as primary packaging of food products, the properties like puncture and tensile strength are significant. The puncture strength is expressed in N/mm as the maximum puncture strength F_{max} divided by the average thickness of the specimen T (Eq. 1.8).

$$PS = \frac{F_{max}}{T} \tag{1.8}$$

Tensile strength is calculated from the maximum tensile strength F_t divided by the product of the average sample thickness T and width W (Eq. 1.9) [31].

$$TS = \frac{F_t}{TW} \tag{1.9}$$

Due to the incorporation of active molecules into polymer matrix, the significant changes in mechanical properties can occur, as the consequence of the sample structure disruption [32].

Surface wettability

Surface properties, as regards the hydrophilicity/hydrophobicity can be determined by the measurement of contact angle between a solid surface and the tangent leading from the contact position of liquid on the substrate (Fig. 9). When a sample hydrophobicity increases, a higher contact angle is obtained [33]. In case that the contact angle approaches zero, the surface is completely wetted by a liquid. If the contact angle is $\geq 90^{\circ}$, the liquid droplet does not spread easily. When the volume of liquid drop is changed, the different contact angles can be obtained. During the liquid is withdrawal, the angle is lower when compared to the situation when the liquid is slowly added. Then, so called *advancing contact angle* θ_A is obtained as the liquid droplet expanding is finished, or the *receding contact angle* θ_R is acquired, when the liquid has just been withdrawn. The values close to 180° are characteristic for so called completely non wetting liquids [27].

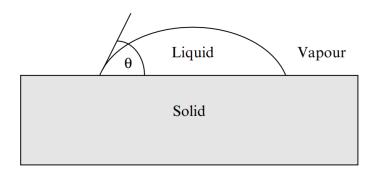


Fig. 9. The contact angle formed by a liquid drop on the solid surface [34]

Contact angles can be measured by different techniques, including sessile drop, captive bubble, or Wilhelmy plate method. However, the right preparation of tested surface is a crucial factor for the successful analysis. Within the author's research, a sessile drop method was applied to get the information on hydrophilicity of polymer surface, which might provide valuable knowledge on e.g. the eventual attachment of active molecules, or the potential antifouling activity useful for many practical applications, such as water treatment. Sessile drop technique utilizes a liquid drop placed on the surface, on which the contact angle is measured. To add or withdraw liquid, for measuring the advancing or receding angle, respectively, a syringe or micropipette is applied. Various types of sophisticated equipment are available at present, such as the video-based contact angle measuring system enabling the analysis of both static and dynamic contact angle and determining the surface free energy of solids and their components, as well as the interfacial tension based on the drop shape analysis [27].

The contact angle depends on the surface tension/energies vectors of individual phase boundaries and can be expressed by the Young equation (1.10):

$$\cos\theta = \frac{\gamma_{SG} - \gamma_{SL}}{\gamma_{LG}} \tag{1.10}$$

Where $cos\theta$ is the contact angle, γ_{SG} , γ_{SL} and γ_{LG} is the surface tension in mN/m at solid/gas boundary, solid/liquid boundary, and liquid/gas boundary, respectively.

A surface energy can be evaluated from the contact angle measurement. Hydrophilic surfaces having lower contact angle exhibit a higher surface energy whereas more hydrophobic substrates have lower water wettability associated with higher contact angles and lower surface energy [35]. Wettability of solid surfaces can be significantly affected by a surfactant adsorbed at the phase

boundaries. In many practical applications, surfactants act as stabilizers, emulsifiers, dispersants or wetting agents.

Testing of antimicrobial properties

Antimicrobial properties of polymer materials can be determined by several methods, depending on the specific application, external factors and expected microbial species that encounter the tested system, since different microorganisms require varying nutritive media and conditions for their growth. The form, shape and geometry of the polymer sample play an important role, too. The antimicrobial efficiency is strongly dependant on the potential and rate of migration of active molecules from the polymer, different interfacial phenomena, as well as the hydrophilicity of the surfaces.

Due to the present problem of an increasing resistance of bacteria to various antibiotics, the methods for evaluating the antibiotic efficiency have been developed. These include a disk diffusion (Kirby Bauer, Stokes test), broth dilution (determination of the minimum inhibition concentration, MIC), diffusion and dilution (E-test) method. During the disk diffusion procedures, the samples are positioned on the agar plates with solid nutritive media containing a microbial suspension of defined concentration. When antibiotics are tested, these are impregnated into sterile filter disk. In case of polymer sample, this is cut into an appropriate size and placed onto the agar plate. Afterwards, the incubation proceeds; time and temperature depend on the type of microbial culture used. After the incubation time, the inhibition zones around the samples are recorded (Fig. 10). Within the Stokes test, an agar plate is divided into three sections containing different microbial cultures [36].

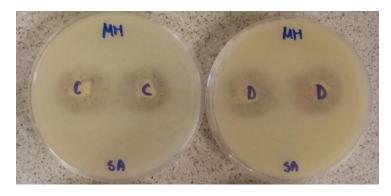


Fig. 10 Inhibition zones around the chitosan samples obtained by an agar diffusion test against Staphylococcus aureus

Analysis of minimum inhibition concentration (MIC) belongs among the most important quantitative testing techniques. A micro-dilution method is usually applied, in which test lakes with equal volume of basic solution and microbial concentration is used for testing various concentrations of antibacterial

agents. After the incubation time, the numbers of colony forming units (CFU) are recorded, either manually or by microscopy. In the E-test, a pre-prepared rectangular strip with a defined concentration of antibiotic solution (of exponential gradient) is placed onto an agar plate. After the test, the MIC value is observed from the scale, from the area, where the ellipse edge intersects the strip [36].

It is known that the antibacterial properties of polymer materials are commonly ensured by the modification with some active molecules. Then, the key factor consists in the ability of these substances to migrate from the matrix in the sufficient level and velocity. The selection of testing methods of such systems is strongly affected by the intended application area. In case when the polymers are going to be used in the food industry, the focus should be put on *Listeria*, *Salmonella* and other indicator microorganisms (e.g. from the family of Enterobactericeae). In the medical applications, so called Methicillin resistant *Staphylococcus aureus*, MRSA, pathogens are investigated [36].

Antifouling activity is another important characteristic that should be considered for surfaces applied in the areas with the excessive tendency to microbial fouling, such as water treatment technologies [36]. Testing of antifouling properties can proceed by different techniques. One of the methods uses the direct measurement of the permeate flux during filtration process. Decrease in the filtration rate usually indicates the membrane fouling. The fouling process can be also analysed by the microscopic techniques if the membranes with sufficient optical transparency are applied [37]. Bacterial adhesion test and comparison of viable bacterial counts in planktonic and sessile form can be also performed to test the antifouling properties [38].

2.2.5 Polymer-surfactant interactions

Mixtures of polymers and surfactants exhibit an interesting behaviour both in solution and at the interface. Thus, they are used in a wide spectrum of practical applications, such as food industry, cosmetics, controlled drug delivery and stabilization of dispersion systems, where they affect rheological properties, stability and surface adsorption [39, 40].

Different kinds of interactions can occur between the polymers and surfactants, including e.g. hydrophobic interactions, or strong electrostatic interactions between charged surfactant head groups and oppositely charged polyelectrolytes [41]. Polymer/surfactant interaction depend on many factors, such as charge density, surfactant ionicity and potential forces between the polymer chain and surfactant monomer or micelle [39].

An interesting phenomenon occurs when the surface tension of the solution containing the surfactant and polymer combination is analysed. In contrast to common surfactant solution (without polymer), the situation is more complex and Gibbs adsorption isotherm is not always applicable below CMC to determine the surface composition. This is due to the fact that the tension discontinuities do not always indicate the appropriate phase changes in the bulk. On the other hand, surface tension measurement still represents one of the most commonly used macroscopic techniques for the investigation of these mixtures.

Figure 11 shows the plot of surface tension vs. surfactant concentration for the case of the weak interactions between the polymer and surfactant in aqueous solution, along with the surface tension behaviour of surfactant solution without polymer. Three obvious points can be identified here, indicated as T₁, T₂ and T₃. The first break, T1, corresponds to the critical aggregation concentration (CAC), at which so called hemimicelles (aggregates of surfactants directly connected with polymer chain) start to form. T2 is related to the area where the bulk polymer is almost saturated with surfactant micelles, thus, the surface tension decreases again until T3 is reached (which is related to critical micelle concentration, CMC) [41].

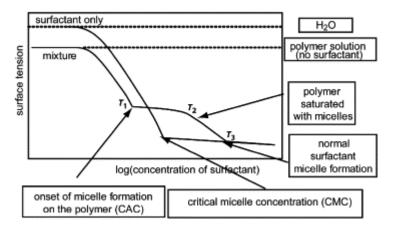


Fig. 11 Surface tension vs. surfactant concentration in the system with and without polymer [41]

Since the surfactant and polymer interact at the surface, an adsorption can occur at significantly lower surfactant concentrations when compared to the solution without polymer. The interactions of less common surfactants and polymers are being studied at the author's workplace. The complexes of nonionic block copolymer from the group of poloxamers (formed by poly(ethylene oxide) and poly(propylene oxide) chains) combined with anionic dioctyl sulfosuccinate were prepared and characterized. The publication of related data is in process.

2.3 Active agents

Various organic or inorganic agents can be used as active agents for the incorporation into polymer matrix, which might then have antioxidant, antimicrobial, antiviral or preservation effects. These substances can be classified according to different criteria, such as the chemical structure, source, spectrum of activity, or function [42].

2.3.1 Organic active agents

Organic antimicrobials act at low concentrations offering a favourable cost/performance ratio for polymers processed at temperatures not exceeding 250 °C. Several substances can be included here, such as antibiotics, vitamins, essential oils, antioxidants, quaternary ammonium compounds etc.

Vitamins can serve as active substances especially for their ability to act as natural antioxidants. Antioxidants are molecules that are able to slow down oxidation mechanisms in food, cosmetics, beverages and pharmaceuticals industry. They can be used as active ingredients and supplements with health benefits, and as stabilizers. Application of antioxidant can be associated with some limitations, such as allergic reactions or unpleasant taste or smell of the product. Additionally, stability of antioxidants might be affected by pH, light, temperature, moisture and oxygen, and can interact with other matrix components, which leads to their degradation and loss of activity. The appropriate encapsulation procedure can solve these issues and increase the resultant product stability [10].

For example, vitamin C (ascorbic acid) is known for various pharmaceutical and biological functions, e.g. it can provide photo protection, scavenge free radicals and generally increase the immunity. To prevent the above-mentioned instability, several approaches of its encapsulation have been studied. Yang et al. have encapsulated vitamin C into a biocompatible layered inorganic material based on hydrated metal oxide. Physico-chemical characteristics, chemical stability along with release profile have been investigated. The prepared system provided sustained vitamin C release and optimal transdermal penetration conditions, which makes it perspective for the application in cosmetics [43]. In the study of Hamadou et al. [44], vitamin C has been encapsulated into marine phospholipids nanoliposomes. A satisfactory stability was proved by testing of the encapsulation efficiency, zeta potential, and index of polydispersity. Vitamin E (α-tocopherol) is known as a strong antioxidant scavenging peroxyl radicals that is capable to protect the lipids from auto-oxidation. Therefore, it is widely applied in healthcare and cosmetics. Due to its light, heat and oxygen sensitivity, and poor solubility, various strategies of incorporation into suitable matrix (solid lipid

nanoparticles, liposomes or nanostructured lipid carriers) have been studied [45]. Teixeira et al. [46] investigated the physical stability od D-α-tocopherol formulated in O/W nanoemulsions based on medium chain triacylglycerols and stabilized by Tween 80 surfactant and soya lecithin hydrolysate as a cosurfactant. The cell viability testing proved a potential of prepared emulsions as drug delivery systems for *in vivo* administration. In another study [47], α-tocopherol was entrapped in Pickering emulsions, in which oil droplets are physically stabilized by colloid particles instead of common emulsifiers. High physical and oxidative stability was proved.

Curcumin has attracted interest of many researchers in recent years. This yellow-orange polyphenol is known for its anti-inflammatory, antioxidant, anticarcinogenic and anticholesterolemic activities. However, it is poorly water soluble, unstable to oxygen, heat, light, enzymes, high pH, and due to rapid metabolism, a rapid systemic elimination occurs. Thus, the various delivery systems have been designed and investigated. Goëlo et al. [48] proposed polysaccharide based carriers prepared by spray drying technique. The release profile was affected by the encapsulating material. From four biopolymers tested (pectin, maltodextrin, inulin, xanthan), the most rapid release was observed in case of inulin. Lycopene is another lipophilic antioxidant that can be also applied as pigment providing a desirable colour to skin [49]. A vesicular delivery system based on ascorbic acid-6-palmitate, cholesterol and dicetyl phosphate was developed for the entrapment and stabilization of lycopene in [50]. Scavenging activity of such encapsulated lycopene was increased when compared to the free lycopene solution.

Essential oils (EO) are volatile aromatic compounds derived usually from plants. Their main applications include fragrances in cosmetic industry; some of them can be used for skin healing and aromatherapy. EO might be incorporated in various formulations to prevent their evaporation or oxidation mechanisms [49]. Within the author's publications, oregano and marjoram (**P8**) and thyme (**P10**) essential oils were used for incorporation into polymer matrices and will be discussed in appropriate sections (Chapter 3.1.4).

Antibiotics can be classified according to different criteria, such as type of action, source, or chemical structure. Based on the spectrum of activity, narrow, medium and wide spectral antibiotics are known. Within the author's works (**P4** and **P6**), three antibiotics from the last mentioned group were tested (ampicillin, minocycline, rifampicin). Antibacterial activity of minocycline/rifampicin coatings was investigated within several studies. The results showed the sufficient effect against Gram positive and Gram negative bacteria, as well as *Candida* species [51]. Nisin, belonging to the group of small-peptide antibiotics, was applied in the papers **P9** and **P12**. This natural

antimicrobial agent is the only bacteriocin allowed for the use in food industry as a preservative [22].

2.3.2 Inorganic active agents

The antimicrobial activity of metal ions, such as silver, copper, or zinc has been widely investigated. Their microbicidal effect can be achieved by affecting the electrochemical potential between the internal and external cell components, or by penetrating into the cells, to compete with other essential ions like magnesium, calcium or potassium, and aggregating with thiol groups of enzymes or proteins. The advantage of inorganic antimicrobial agents consists in the negligible impact on the processing mechanisms and the potential use for the polymers processed at high temperatures. They are highly recommended for the applications requiring long-lasting antimicrobial activity and the risk of bacterial resistance is reduced in comparison to other antibacterial agents (such as antibiotics) [52].

Within the author's publications (**P1**, **P2**, **P3**, **P5**, **P7**), silver and zinc ions were investigated for incorporation into polymer matrices by solvent casting or thermoplastic compounding. Discussion on these active agents is included in the appropriate chapters of the thesis (3.1.1 to 3.1.3).

3. DISCUSSION ON PAPERS

Within the following chapters, the main author's research subjects will be overviewed, classified into the sections according to the form of the polymer system, i.e. films, thin layers and nanofibrous membranes.

3.1 Polymer films

Polymer films represent perspective carriers for embedding various active molecules that can serve as "smart" coatings and packaging for preserving foods, or as wound dressings. These systems can control the gas or moisture permeability, prevent oxidation mechanisms, limit the biofilm formation and thus prolong the product shelf life [53]. Synthetic and/or natural polymers might be used for the preparation of such systems.

Synthetic polymers gained a high portion of attention in many industrial fields, especially due to a variability of physical and chemical properties that can be achieved, based on the character of monomer units, type of polymerization reaction and potential formation of copolymers with different components [54]. In bioactive systems, hydrolytically degradable polymers are often applied, whereas the process of enzymatic degradation is preferred for biopolymers.

With respect to author's publications, the following text is divided into the chapters according to the processing technique (the melt compounding and solvent casting), and the base polymer used as the potential carrier of active substance. Several types of polymers were investigated, such as vinyl polymers, poly(vinyl alcohol) and poly(vinyl chloride), as the representatives of synthetic materials, and chitosan from the group of biopolymers.

3.1.1 Poly(vinyl alcohol) based films prepared by solvent casting

Poly(vinyl alcohol) (PVA) is a synthetic biocompatible, biodegradable, water-soluble polymer, properties of which are driven by the preparation method and hydrolysis degree [55, 56]. It can be applied in different commercial sectors, e.g. for the production of artificial skin replacements, contact lenses, food packaging systems, and/or carriers for the drug release. Combinations of PVA and antibacterial agents, such as zinc or silver, have been studied within the authors' publications (**P1**, **P2**, **P3**). Active compounds were encapsulated into polymer matrix by the casting technique, which represents a promising solution in the development of antibacterial polymer system with a longer-term effect fulfilling the high quality requirements [57, 58, 59]. Films with uniform thickness and low haze can be obtained in this way [60].

Active multilayer materials have been investigated in the work of Tampau et al. [53], in which the polymer solution with active compound is placed on an appropriate polymer support. The efficiency of such prepared multilayers was significantly affected by the wettability of solution and the spreadability on the

supporting layer. Therefore, the contact angle of the polymer-solution system and surface tension of the applied solutions play a crucial role, too.

Silver compounds have been known as broad-spectrum antibacterial agents for many years. Their activity is attributed to the interaction of Ag⁺ ions with bacterial DNA, which leads to the loss of its replication ability. In addition, the effect of Ag⁺ ions on the membrane-bound enzymes has been reported. Silver nitrate (AgNO₃), known for its toxicity to bacteria, can serve as the source of silver ions. Poly(vinyl alcohol) films enriched with silver nitrate were prepared by the solvent casting in **P1**. The thermal and spectral analysis proved the presence of the silver particles in PVA matrix. Surprisingly, the mechanical properties were positively affected by the addition of active agent up to 1 wt.% and the tensile strain was even enhanced over the whole tested concentration range of Ag (0.5 to 9 wt.%). Prepared systems proved a high activity against both tested Gram negative (*E. coli*) and Gram positive (*S. aureus*) microorganisms, which makes them useful in different applications including the protecting packaging.

Poly(vinyl alcohol) carrier was also used in the papers P2 and P3, however, zinc ions were selected as active substance for the encapsulation. It is known that its antibacterial activity consists in the formation of zinc ions in aqueous environment that bind the cell membranes, thus prolonging the lag phase. Specifically, zinc nitrate (Zn(NO₃)₂), as one on the most frequently reported zinc source, was applied in P2 in the concentrations recalculated for Zn from 0.5 to 9 wt.%. Mechanical testing proved an increase of E modulus up to 3 wt.% Zn content, after which a gradual decrease was observed, with further addition of modifier. A similar trend was shown for tensile strength, whereas tensile strain was decreasing within the whole concentration range of Zn. The results revealed that the low zinc concentrations (0.5, 1 and 3 wt.%) enhanced the toughness. On the other hand, due to a higher zinc content, the mechanical properties were generally weakened. Differential scanning calorimetry proved an initial increase (up to 72 °C for 3 wt.% Zn) of the glass transition temperature (Tg) at the lower modifier concentrations followed by the further decrease (up to 55°C for 9 wt.% of Zn). These observations indicated that the changes in the polymeric chains mobility occurred. The structural analysis by Fourier transform infrared spectroscopy (ATR-FTIR) of the PVA/Zn films proved a uniform distribution of zinc nitrate in the polymer matrix. Antibacterial tests confirmed the inhibition activity of prepared samples against the selected bacteria, even from the lowest tested zinc concentration (0.5 wt.%) against Staphylococcus aureus.

PVA matrix enriched with zinc sulphate (ZnSO₄) was investigated in the publication **P3**. The resultant physico-chemical and antibacterial properties, as well as the effect of PVA hydrolysis degree was investigated. The type of PVA proved to have a significant impact on the distribution of zinc sulphate within the PVA matrix, and mechanical and antibacterial properties, too. The films based on the fully hydrolysed polymer exhibited notable increase in Young's modulus and tensile strength. On the other hand, these samples revealed a hindered release of

active agent from the matrix, probably due to the different solubility of partially and fully hydrolysed PVA. From the microorganisms tested, Gram positive *Staphylococcus aureus* proved to be the most sensitive, whereas Gram negative bacteria showed higher resistance to the tested films (*Pseudomonas aeruginosa* exhibited zero inhibition zones for the fully hydrolysed PVA samples). Beside the common agar diffusion test, a dilution and spread plate technique was applied, which is considered as the highly sensitive and accurate method enabling to evaluate the activity in liquid medium. The higher sensitivity of Gram positive bacteria, as well as the effect of PVA hydrolysis degree, was also proved by this procedure, due to the obtained parameters of the effectiveness of antibacterial activity (*EAA*) and the efficiency constant (-k), which informs on the antibacterial efficiency of the selected agent under specific conditions.

3.1.2 Poly(vinyl chloride) based films prepared by thermoplastic compounding

Poly(vinyl chloride) (PVC) molecule contains ethylene backbone with covalently bonded chlorine. It is one of the most frequently applied thermoplastics with versatile properties, good mechanical strength, chemical resistance and inertness to biological fluids [61]. Modification of medical grade PVC with organic/inorganic active substances was investigated in the publications (**P5**, **P6**).

Thermoplastic compounding or melt blending technique has been extensively applied for the preparation of polymers with uniformly distributed antimicrobial agents enabling a long-term activity. Various antimicrobial agents can be incorporated into polymer matrix. From the group of inorganic substances, silver and its derivatives has been intensively studied due to its well-known antimicrobial activity based on the silver cation Ag⁺ that strongly binds to electron donor parts of biomolecules containing sulphur, nitrogen, or oxygen. The advantage of silver-based polymer composites prepared by thermoplastic compounding consists not only in the high antimicrobial effect, stability and non-toxicity of silver ions to human cells, but also in the great temperature processibility [62].

In the publication **P5**, a medical-grade PVC was used for thermoplastic compounding with silver nitrate (recalculated to Ag content from 0.5 to 5 wt.%) in a Brabender Plasti-corder kneader. The process was carried out at 160°C and 30 rpm for 10 min, after which the samples were compression molded at 160°C for 5 minutes in the manual press to obtain the thin films. This polymer substrate was selected as a carrier for its good compatibility with a number of additives, its mechanical properties and suitable economical characteristics. The aim of the author's work was to study the effect of modification on the final physicochemical and antimicrobial properties. Incorporation of a modifier led to a decrease of tensile characteristics, probably due to an infracted structure caused by silver aggregates. In comparison to the above mentioned results from **P1**, where the improvement of mechanical properties (Young's modulus, tensile

strength) was observed up to 1 wt.% of silver concentration in PVA, it can be assumed that the polymer matrix, as well as the processing method, play a crucial role in the final properties of the antibacterial polymer system. Nevertheless, the obtained nonhomogeneous structure of PVC/Ag samples did not negatively affect the antibacterial action. All tested bacterial strains including both the Gram positive and Gram negative ones (*Klebsiella pneumonia*, *Escherichia coli*, *Pseudomonas aeruginosa*, *Staphylococcus aureus*) proved to be sensitive to prepared PVC-silver composite, even at the lowest filler concentration (0.5 wt.%).

The method of thermoplastic compounding of PVC was also applied in the further study (**P6**), however, three types of antibiotics were incorporated (0.1 to 1 wt.%) in the polymer matrix to obtain the system with antibacterial properties that could be potentially applied in a big-scale production. The active substances (sodium ampicillin, minocycline, rifampicin) were selected with regard to their activity spectrum, functional groups (Fig. 12) and the thermal stability, too. Regarding the effect of the PVC modification with antibiotics, a key role of their chemical structure as well as physico-chemical properties was confirmed. X-ray diffraction measurement revealed that processing conditions (160°C, 10 minutes) did not have any negative impact on the structural properties of used antibiotics.

Fig. 12 Structures of sodium ampicillin, minocycline, rifampicin [63]

It is known that mechanical characteristics of polymer materials have a significant impact on the potential applicability of the product. Although a decrease in tensile strength and strain was observed in case of all antibiotics, the effect on E modulus was rather negligible.

Considering the antibacterial properties, all tested samples inhibited the growth of Gram positive *Staphylococcus aureus*, whereas Gram negative *Escherichia coli* proved to be more resistant against lower antibiotic concentrations; and completely inert to rifampicin samples. Microscopic evaluation confirmed a

higher degree of incorporation, which definitely let to its limited migration from polymer matrix.

This statement was supported by the kinetic study that proved an impact of different molecular structures of active agents on their release from the matrix. While in case of ampicillin and minocycline a burst effect was observed within the initial phase of testing, rifampicin release was slower. This different behaviour can be confirmed by the appropriate constants shown in Table 1, where the highest concentrations of all three types of antibiotics (1 wt.%) were compared.

Table 1 Constants achieved by the release studies of antibiotics from PVC systems

Antibiotic	Distilled water			Physiological solution		
(1 wt.%)	C _{max} (µ/g)	-k (1/h)	\mathbb{R}^2	C _{max} (µ/g)	-k (1/h)	\mathbb{R}^2
ampicillin	2006	123	0.99	2359	0.26	0.99
minocycline	720	0.02	0.99	110	1.00	0.99
rifampicin	152	0.04	0.97	206	0.02	0.99

It is necessary to state that the amount of active molecule released into the environment is strongly affected by the nature of the polymer matrix (hydrophobicity) and the type of entrapment. It is clear that the released concentrations differ depending on the type of antibiotic used; from which the maximum released amount (C_{max}) was observed for ampicillin. When the initial antibiotic loading is considered, it takes only 0.2 wt%. However, it is known that the release course is controlled not only by the diffusion, but also by the area of the interface related to the polymer surface/surrounding environment. Therefore, in practical applications, this parameter will be predominant.

3.1.3 Poly(vinyl chloride) based films prepared by solvent casting

To compare the effect of the processing technique on the character of the films, a medical-grade poly(vinyl chloride) based films enriched with sodium ampicillin prepared by the solvent casting were investigated in the author's publication **P4**. A significant impact of applied solvent (cyclohexanone, CYH, or N,N-dimethylformamide, DMF) was confirmed. Tensile properties were enhanced in case the film was cast from CYH, whereas the latter mentioned solvent caused an opposite effect.

The effect of different nature of used solvents on the film properties was proved even in the morphological study. Whereas CYH-based films were smooth with uniform filler distribution, the DMF sample contained clusters of ampicillin on the surface (Fig. 13). On the other hand, these samples showed a more significant inhibition effect against all tested bacteria.

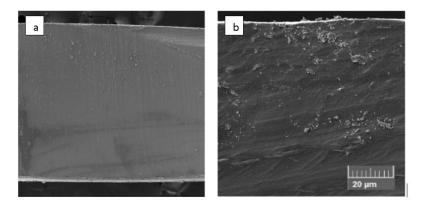


Fig. 13 SEM microghraphs of (a) PVC/CYH/5 wt.% ampicilin and (b) PVC/DMF/5 wt.% ampicillin

A first-order kinetic was successfully applied to release experiment; slower release of ampicilin was observed from cyclohexanone samples, whereas a burst effect was shown in dimethylformamide solvent. This different behaviour was evidenced by the constants obtained from the release studies, when the maximum theoretical antibiotic concentration released from 1 g of the sample (C_{MAX} in $\mu g/g$) was two orders of magnitude higher when compared CYH (41 $\mu g/g$) and DMF (1026 $\mu g/g$) sample modified with 1 wt.% of sodium ampicillin.

Beside the release of ampicillin into water, the experiment in the physiological solution was also carried out to evaluate the effect of ionic strength. It was shown that substantially higher antibiotic content was released into physiological solution, both in case of cyclohexanone and dimethylformamid, probably due to the affinity if ampicillin to isotonic solutions.

The casting technique using the same solvents, CYH and DMF, was applied in the author's publication **P7**, where the addition of silver nitrate (recalculated for the silver content from 0.5 to 5 wt.%) into PVC carrier, concretely the effect on the final structural, mechanical and antimicrobial properties, was investigated,. Prepared samples showed a significant antimicrobial activity even at the lowest tested silver concentration (0.5 wt.%). Antimicrobial effect remained almost unaltered even after 24h immersion in physiological solution. Surface analysis by X-ray photoelectron spectroscopy confirmed that PVC based prepared samples contained the silver mostly in Ag⁰ form and the immersion in the physiological solution resulted in the transformation to Ag⁺, due to the presence of AgCl. The incorporation of silver nitrate had a negligible impact on mechanical properties. The interesting results came from the comparison of the tensile characteristics

measurement of the PVC samples prepared in CYH and/or DMF, since no significant differences were observed.

3.1.4 Natural polymer/surfactant based films prepared by solvent casting

With increasing demands on environmentally friendly, well degradable materials, a number of present works are oriented more toward natural polymers derived from plant or animal sources. Polysaccharides and proteins are the common representatives of natural polymers that can be applied as suitable carrier materials for various active agents [64].

Chitosan belongs to polysaccharides derived mostly from crustacean cells. From the chemical point of view, it is linear copolymer based on D-glucosamine and N-acetyl-D-glucosamine, linked by β - $(1\rightarrow 4)$ bonds. Chitosan is biocompatible, biodegradable, nontoxic polymer that even possess antimicrobial properties, which predetermines it for application in many industrial sectors, such as medical, food or water treatment industry. During the last decade, chitosan and its derivatives were applied either in form of gels, coatings, food packaging or wound dressings. A solvent casting technique with following evaporation of solvent represents the standard method for a film preparation. Several aspects, as low water vapour permeability, high mechanical strength, sufficient stability and antimicrobial properties, have to be considered [65]. Although chitosan itself offers antimicrobial activity due to interacting of its positively charged amino groups with negative bacterial membranes, this effect can substantially decrease when it is in the form of film. Following this fact, different active substances have been tested for incorporation into chitosan films [66, 67].

Essential oils and their main components belong to attractive candidates, due to their interesting biological activity. Their structure and actually their functional groups play a crucial role in resultant antimicrobial properties. In this respect, phenolic and terpenoic compounds proved to be highly effective [68]. Several studies have been dealt with the combination of chitosan and different essential oils, such as lemon, rosemary, basil, cinnamon or clove [69].

The chitosan films were investigated in the papers **P8** and **P10** that were aimed at the study of mutual interactions between polymer, active molecule and surfactant. The author's paper **P8** is devoted to a complex study of oregano and marjoram essential oils incorporated into chitosan matrix, regarding both film forming solutions and resulting films. The principle consisted in the addition of a third component, non-ionic surfactant Tween 80, to ensure the enhanced dispersibility and stability of chitosan samples.

Since the properties of individual essential oils are significantly affected by the specific functional groups present in the molecule, the chromatography analysis is recommended to obtain the appropriate data on the molecular structure. Although both oregano and marjoram comes from the same *Origanum* genus, gas chromatography of applied essential oils revealed a different composition of

active components that affected physico-chemical and antimicrobial properties of prepared solutions and films. Oregano oil included carvacrol as a predominant substance, which is known for its strong antioxidant and antimicrobial properties. On the other hand, an alcohol terpinen-4-ol was found as the main constituent in marjoram essential oil. These findings reflected in the antimicrobial properties. While oregano samples proved a high antimicrobial efficiency against all tested strains that was even enhanced after one week in some cases, marjoram films did not show any significant inhibition effect, probably due to a more compact structure limiting diffusion of active agent into surrounding. The addition of nonionic surfactant affected the properties of film forming solutions when the particle size significantly decreased with an increasing Tween 80 concentration, regardless the essential oil used. Moreover, a good dispersion stability of both oregano and marjoram film forming solutions was achieved, based on the zeta potential values, which exceeded +30 mV in all samples. For the polymer films intended for use in packaging applications, the barrier properties play a substantial role. Water vapour permeability (WVP) is the parameter indicating the duration of water vapour transmission through the unit area of tested material, evaluating the difference between the unit vapour pressure between two surfaces (under specific temperature and humidity). The results showed that modification of chitosan with essential oil/surfactant mixture led to WVP decrease (e.g. from 16.5 \times 10⁻³ g/Pa.h.m² for pure chitosan to 7.8 \times 10⁻³ g/Pa.h.m² for chitosan sample with 5 wt.% oregano/Tween 80) and thus, to enhancement of barrier properties. Based on the findings in this publication, chitosan/oregano/Tween 80 films have the potential for the efficient bioactive coatings.

The effects of thyme oil incorporation into chitosan matrix were investigated in the publication **P10**, which was predominantly focused on the type of stabilizer used. Three types of non-ionic surface active agents with different hydrophilic-lipophilic balance (HLB) values belonging to the group of ethoxylated sorbitan esters (polysorbates, Tweens) were selected. A more homogeneous structure was obtained with the emulsifiers of higher HLB (Tween 80 with HLB 15 provided the most stable system). Due to the modification of chitosan with Tween/thyme oil combinations, stable polymer films with sufficient antimicrobial and enhanced water barrier properties were prepared by cost-effective and environmentally favourable way (Fig. 14). In addition, the final physico-chemical properties proved to be tuneable by the surfactant/stabilizer selection.



Fig. 14 Appearance of the chitosan films a) without and b) with thyme essential oil/Tween 80

The interesting results were obtained by the morphological analysis of the films. Fig. 15 showing the SEM photographs of the chitosan films reveals the different structure depending on the added active substance. Samples with thyme and oregano essential oil contain bigger particles (Fig. 15 a, b) compared to the film with marjoram oil (Fig. 15 c). This corresponds to the particle size measurement of the appropriate film forming solutions, where the latter mentioned sample exhibited one order of magnitude smaller particles. Moreover, a more compact structure was observed in marjoram films, probably due to more hydrophilic components. On the other hand, this arrangement hindered the diffusion of active agent from the chitosan matrix, which resulted in a weaker antimicrobial activity in comparison to thyme and oregano oil.

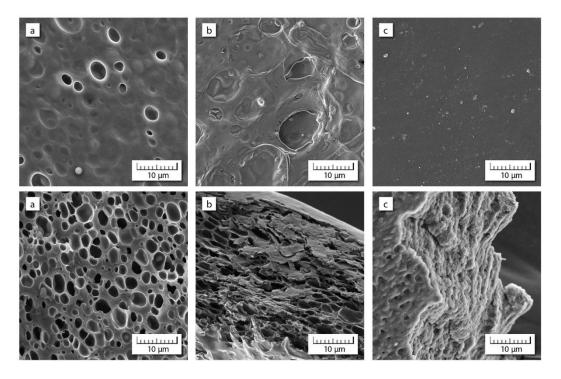


Fig. 15 SEM micrographs of chitosan films with Tween 80 and 2 % essential oil: a) thyme, b) oregano, c) marjoram oil: up (surface), down (cross section)

To compare the effect of polymer matrix, as well as the selected incorporated active molecule, the non-published results of the ongoing research at the author's workplace dealing with zein films are shortly mentioned. Zein belongs to the class of plant proteins known as prolamines that occur specifically in cereals. It is insoluble in water and can dissolve only in the presence of alcohol or high concentrations of anionic detergents. Zein is known for its valuable potential for the formation of films that provide optimum moisture and oxygen barrier properties. Due to this fact, different antibacterial and antioxidant agents, such as salicylic acid and its derivatives, nisin, thymol, as well as cationic surfactant lauroyl arginate, were incorporated in zein matrix to achieve the bioactive protein based biodegradable films [70–75]. Within the author's research, zein films enriched with the combinations of essential oils (thyme or oregano) with monoacylglycerol of lauric acid (monolaurin, MAG 12) were prepared by the solvent casting technique with the aim to evaluate the potential synergic effects of these two types of active agents. Comparison of the results obtained by the SEM analysis for chitosan and zein (Fig. 15 and Fig. 16) reveals the significant difference in morphology when the crystals of various shapes and size and the key role of added monolaurin was monitored in the zein samples.

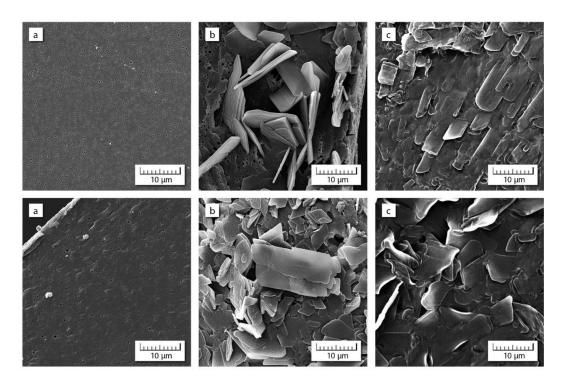


Fig. 16 SEM micrographs of zein films with: a) 4%MAG12/2% thyme oil, b) 5%MAG12/2% oregano oil, up (surface), down (cross section)

3.2 Plasma polymer thin layers

Several approaches are utilized to modify polymer substrate surface properties in order to enhance the consequent attachment of active molecules. Plasma technologies represent clean, environmentally favourable methods of control the chemical and mechanical properties of polymers. The advantage consists in the fact that the material surface can be modified without a significant alteration of bulk properties, due to which original mechanical characteristics are preserved. Ultra-thin films with controllable properties can be obtained by plasma treatment. Moreover, speed, practical scalability of the process and no need of solvents application make this technique popular for the production of active substances carriers [76].

Different plasma treatment methods can be used, such as low temperature short term irradiation for material surface modification, or high intensity plasma treatment preferred more for surface sterilization. For the preparation of thin films, plasma-enhanced chemical vapour deposition (PECVD) can be applied. The film properties, such as density, thickness and roughness depend on various factors, including sputtered precursor, flow rate and/or type of gas in a plasma chamber. Within the study of controlled active agent release from polymer materials modified by plasma treatment, majority of experiments are carried out in air, however, atmosphere or other gases (nitrogen, argon) can be also used [76].

A number of systems for controlled release of active molecules prepared by plasma treatment have been investigated, such as films, microparticles, electrospun membranes or tablets [77–79]. It is known that release of active agents from polymeric materials generally occurs via diffusion. However, degradation process can play an important role, too, especially for easily degradable systems. In this case, a surface wettability has to be considered as the factor affecting a degradation rate as well as a diffusion coefficient of an incorporated drug and consequently its release kinetics. In this context, plasma treatment can be applied as an efficient tool for modifying the hydrophilicity of polymer materials surfaces [76].

The effect of plasma pre-treatment and a crosslinking degree on the poly(vinyl alcohol) (PVA) based films was investigated in the publication **P9** in order to evaluate their potential for carriers of bioactive molecule, nisin. PVA is known for its biodegradability, good mechanical properties, large swelling capacity and low cost. This polymer contains a number of potential functional groups enhancing the adhesion of various active or preserving agents [80]. Surface plasma dielectric coplanar surface barrier discharge (DCSBD) was used to modify surface hydrophilicity of PVA film so that an enhanced adhesion of nisin is achieved. This technique uses atmospheric plasma surface activation operating in a non-thermal, uniform, glow plasma regime and can be used even for large area surface modifications [81].

The results proved that both crosslinking degree and plasma discharge power are crucial for PVA layer/nisin layer stability. A strong nisin attachment was achieved in the films with 10 and 20% crosslinking degree. In these samples, a significant increase in wettability by the plasma treatment was observed. Consequently, the higher total surface energy was monitored, which enhanced a nisin adhesion. Data from release kinetics study revealed that the slowest nisin release was obtained at 20% crosslinking degree after the plasma treatment. Based also on the results of antimicrobial assay, this sample was proved as an efficient and stable system with controllable release conditions.

In the publication **P11**, poly(lactic acid) (PLA) based thin films were prepared by a plasma assisted vacuum thermal deposition (PAVTD) to study the effect of varying radio frequency discharge power on chemical composition, structural properties and hydrolysis rate of prepared thin films. PAVTD technique uses oligomers released during low-pressure thermal decomposition of bulk polymer precursor to form the thin film either directly or after plasma re-polymerization. Due to this fact, various properties of the film can be controlled. Poly(lactic acid) is a synthetic biodegradable polymer widely applied in biomedical field, as well as in food packaging. Its degradation mechanism is based on hydrolysis of ester bonds [82]. High performance liquid chromatography with UV detection (HPLC-UV) was used for determination of lactic acid released into water as a measure of hydrolysis reaction. It was shown that a faster hydrolysis proceeded in the films prepared at lower power, whereas higher effective power resulted in a slower hydrolysis rate. These samples provided a more cross-linked structure although the PLA-like groups were preserved. This was also proved by structural analysis (ATR-FTIR spectroscopy), where prepared PLA plasma films revealed no significant changes when compared to original PLA polymer.

The PAVTD method was also applied in the publication **P12** to obtain films based on poly(ethylene oxide) (PEO) precursors. The aim of the study was to investigate the effect of molecular weight (Mw) of precursor on the physical properties of final system, and the influence of plasma power on the biomolecule permeability conditions. The Mw of the precursors showed a negligible effect on the film properties while the process conditions (presence/absence of plasma) had stronger influence. The rate of thickness loss after immersion in water was found to be tuneable in this way. These films were used as cover and permeation layers for bacteriocin nisin dispersed in PVA carrier. Controlled rate of release of biologically active nisin molecules into water was achieved.

3.3 Polymer nanofibrous membranes

The electrospinning process is another method that can be used for encapsulation of active molecules into polymer matrix. This technique is based on the application of electric charge enabling the production of nanofibers from polymer solutions or melts. When polymer precursor is placed at the pipette tip, the high voltage is generated that leads to the formation of Taylor cones at the polymer material surface [83, 84]. Charged polymer jets pass from the tip to the grounded collector that is accompanied by a reduction of their diameters (Fig. 17).

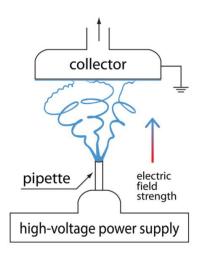


Fig. 17 Scheme of an electrospinning apparatus [83]

The resultant polymer membranes are defined as solid sheets porous enough to enable the passage of small particles that can be applied for permeability of gases, vapours and liquids. Incorporation of organic or inorganic substances have been recently studied to obtain the materials with enhanced magnetic properties, increased hydrophobicity, and antibacterial or antifouling activity [85, 86]. Antibacterial nanofibrous membranes find applications in many industrial fields, such as medical and healthcare, food preservation, protecting of textile materials, or water treatment [87–90]. The active agents, which are most widely utilized for the preparation of antibacterial nanofibrous membranes, include silver nanoparticles, antibiotics, quaternary ammonium salts, or essential oils [91–93]. Materials with antifouling properties gain an increasing interest in the applications, where biofouling of polymer membranes leading to the biofilm formation represent a serious obstacle. Antifouling surfaces prevent the accumulation of biofilms on their surfaces by either affecting the structure of microbial biofilm (due to the steric repulsion or nanoscale rough topography) or repelling microorganisms [94, 95]. Recent research focus on novel strategies bringing about new antibacterial and antifouling surfaces that are safe and do not promote bacterial resistance at the same time.

Antifouling membranes based on poly(vinyl butyral) and monocaprin (MAG 10) were prepared and characterized in **P13**. Poly(vinyl butyral) (PVB) is frequently applied in the electrospinning process to enhance the spinnability of polymer solutions [96]. Monocaprin exhibits antibacterial properties against both Gram positive and Gram negative bacteria, due to which it interferes with

bacterial cell membranes that might lead to the inhibition of cell metabolism or a cell lysis. Already Kabara [97] revealed some general statements concerning the antimicrobial activities of lipid and their derivatives. Regarding monoacylglycerols, those of medium-chain saturated fatty acids proved to be more effective than the free acids. On the other hand, di- and triacylglycerols do not show a significant antimicrobial activity. Antibacterial properties of shortchain saturated fatty acids (from 6 to 10 carbons), where monocaprin is classified, are associated with the dissociation degree, and the higher activity was proved at pH 6.5. MAGs can be utilized in the pharmaceutical industry, or in the protection of textile materials. Since monoacylglycerols have a specific amphiphilic structure, they are applicable as surfactants in the food industry [98]. The positive effects of surfactants on the character of polymer solutions applied for electrospinning was even shown in some of previous studies [99].

Nanofibrous membranes were prepared from various PVB concentrated solutions (8, 10, and 12 wt%) with different amounts of monocaprin (from 0.5 to 3 wt%). Increasing the concentration of monoacygleerol resulted in different shape of the nanofibers, probably due to the fact that MAG 10 stayed embedded in the inner fiber structure. From this reason, the diffusion from the sample was hindered, as well as the antibacterial effect tested by the agar-diffusion method. On the other hand, a good antifouling activity of nanofibrous membranes was shown, when bacterial viable counts in planktonic vs. sessile form was investigated. Simultaneously, the level of wettability was heightened, which is favourable in the development of antifouling surfaces, too. The prepared nanofibrous membranes showed the potential as antifouling filtration membrane in the processes of waste water treatment. Due to a variety of factors affecting the physico-chemical and antibacterial properties, the further research will focus on the investigation of the molecular structure of applied active agent (monoacylglycerols with different chain length will be tested) and mutual interactions with the polymer.

4. CONTRIBUTION TO SCIENCE/PRACTICE

The habilitation thesis summarises the work related to the research on formulation and preparation of active polymer systems, which was carried out from 2008 until present. Within the author's works, the various preparation techniques from the thermoplastic compounding through the solvent casting to electrospinning or the plasma deposition processes have been used. Similarly, the attention was paid to careful selection of antimicrobial agents, of which inorganic (based on silver or zinc ions) and/or organic (based on antibiotics and phenol) compounds were investigated.

The papers **P1 to P7**, published within the years 2008 to 2013, are dealing with the utilization of synthetic polymers, poly(vinyl chloride) and poly(vinyl alcohol), as carriers for selected organic and/or inorganic antimicrobial substances were incorporated by the melt blending or solvent casting technique. The main contribution of these works to science and practice is the successful preparation of the highly efficient and applicable antibacterial polymer systems by using the facile and economically favourable methods.

Along with an increasing trend in searching of environmentally more friendly materials, the attention was directed towards natural polymer matrices based primarily on chitosan, organic active agent and a surfactant, to achieve a better incorporation into polymer matrix (publications **P8** and **P10**). This research unambiguously documented that it is possible to prepare safe, biodegradable, biocompatible and nontoxic films that could be utilized for preserving and protecting the food or cosmetic products from contamination.

The papers **P9**, **P11**, **P12** (2018–2020) were dealing with the plasma deposition used for the modification of the polymer substrate to study the effect on the surface, structural properties and degradability. The interesting pieces of knowledge on the plasma treatment came out of the study on poly(lactide). Here the original polymer structure was well preserved after the plasma deposition, while other properties, including the degradability, proved to be tuneable, depending on the deposition conditions.

The remarkable outcomes with high application potential were provided in the last study **P13** dealing with the novel nanofibrous membranes based on poly(vinyl butyral) and monocaprin. The modification with this non-ionic amphiphilic substance significantly enhanced the antifouling properties of the resulting membranes, which is highly required in the materials applied in the water treatment processes.

The main contribution of the work to the science lies in the increasing the knowledge on the key approaches for incorporation of active agents into polymer and biopolymer matrices, leading to the design of an efficient active polymer based carrier systems. From the scientific point of view, the revealing mutual interactions between polymer matrix, and active or stabilizing agent is crucial, resulting in the deeper understanding of phenomena occurring at surfaces and

interfaces of the dispersion polymer systems. The contribution to practice then consists in the development of functional polymer-based materials providing antimicrobial, and/or antifouling properties, with controllable characteristics, such as degradability, stability and/or release of antimicrobial molecules. These materials show the potential to prevent the bacterial colonization, which is one of the major problems in food, cosmetic or medical industry. The efficient suppression of the bacterial growth favourably affects not only public health, but also environment and economy.

CONCLUSIONS

Various strategies how to incorporate the active substances into polymer matrices were investigated to evaluate the effect of such modification on the final antimicrobial, physico-chemical and surface properties of the products. The work documented successful preparation of the active polymer systems that can help to reduce and even eliminate the microbial contamination affecting a broad range of industrial and human activities. Following the results of the papers dealing with the solvent casting technique, it was proved, that this relatively simple and lowcost method can be applied for production of the efficient antimicrobial polymer systems. In case of poly(vinyl chloride) based films, the crucial role of the solvents used for polymer dissolution prior to casting was observed. Within the current author's research and educational activities, the incorporation of active and/or stabilization substances with the surface activity and their interactions with the polymers have been studied. The key role of the surfactant type and its hydrophilic-lipophilic balance was revealed in the papers investigating the chitosan-based solutions and films, especially in the analysis of their surface and morphological properties. The studies devoted to the plasma-treated polymer thin films showed that their degradation mechanisms as well as the potential to release selected active molecules could be controlled via the applied plasma power. Beside the mentioned techniques, the electrospinning process was successfully utilized for the preparation of the poly(vinyl butyral) membranes with non-ionic surfactant monocaprin, in which the antifouling properties were observed.

To sum up, the introduced polymer-based systems have a good potential to serve as the carriers of different active molecules that could be efficiently used in several practical above-mentioned applications.

REFERENCES

- 1. NARANG, A.S., DELMARRE, D., GAO, D. Stable drug encapsulation in micelles and microemulsions. *International Journal of Pharmaceutics*. 2007, **345**(1), pp. 9-25. doi:10.1016/j.ijpharm.2007.08.057.
- 2. MYERS, D. *Surfactant science and technology*. 3rd ed. Hoboken, New Jersey: John Willey & Sons, Inc., ©2006, ISBN 9780471746072.
- 3. MALMSTEN, M. Surfactants and polymers in drug delivery. New York: Marcel Dekker, Inc., ©2002, ISBN 0-8247-0804-0.
- 4. GARCIA, M.C., ALOISIO, C., ONNAINTY, R., ULLIO-GAMBOA, G. Self-assembled nanomaterials In: Nanobiomaterials: nanostructured materials for biomedical applications. The Netherlands: Elsevier Ltd.: 2018, pp.41-94.
- 5. CHAUDHARI, S. P., PATIL, J. R. Study of block copolymer micelles as vehicles for hydrophobic drug Lamotrigine. *Indian Journal of Pharmaceutical Education and Research*. 2014, **48**(Suppl), pp. 56-66. doi:10.5530/IJPER.48.4S.8.
- 6. PETRŽELKOVÁ, Markéta. Polymeric micelles as carriers of active substances [online]. Zlín, 2020 [viewed 2020-07-11]. Diploma work. Tomas Bata University in Zlin, Faculty of Technology, Department of Lipids, Surfactants and Cosmetics Technology. Supervisor Ing. Jana Sedlaříková, Ph.D. Available from IS/STAG UTB portal.
- 7. FANUN, M. Microemulsions: properties and applications. Boca Raton: CRC Press©2009. In: Surfactant science series, v. 144. ISBN 1420089595.
- 8. CHELLAPA, P., ARIFIN, F. D., EID, A. M. et al. Nanoemulsion for cosmetic application. *European Journal of Biomedical and Pharmaceutical Sciences*. 2016, **3**(7), pp. 8-11.
- 9. TADROS, F. T. *Emulsion formation and stability*. Wiley-VCH Verlag GmbH&Co. KGaA, ©2013 ISBN: 978-3-527-31991-6.
- 10.COSTA, R., SANTOS, L. Delivery systems for cosmetics from manufacturing to the skin of natural antioxidants. *Powder Technology*. 2017, **322**(), pp. 402-416. doi:10.1016/J.POWTEC.2017.07.086.
- 11.LAOUINI, A., JAAFAR-MALLEJ, C., LIMAYEM-BLOUZA, I. et al. Preparation, characterization and applications of liposomes: State of the art. *Colloid Science and Biotechnology*. 2012, **1**(2), pp. 147-168.
- 12.AKBARZADEH, A., REZAEI-SADABADY, R., DAVARAN, S. Liposome: classification, preparation, and applications. *Nanoscale Research Letters*. 2013, **8**(1), pp. 102. doi: 10.1186/1556-276X-8-102.
- 13.MARIANECCI, C., Di MARZIO, L., RINALDI, F. Niosomes from 80s to present: the state of the art. *Advances in Colloid and Interface Science*. 2014, **205**(), pp. 187-206. doi: 10.1016/j.cis.2013.11.018.

- 14.CORTESI, R., ROMAGNOLI, R., DRECHSLER, M. et al. Liposomesand ethosomes-associated distamycins: a comparative study. *Journal of Liposome Research*. 2010, **20**(4), pp. 277-285. doi: 10.3109/08982100903443057.
- 15.DOOST, A. S., NASRABADI, M. N. KASSOZI, V. et al. Recent advances in food colloidal delivery systems for essential oils and their main components. *Trends in Food Science & Technology*. 2020, **99**, pp. 474-486. doi: 10.1016/j.tifs.2020.03.037.
- 16.KENAWY, E., WORLEY, S. D., BROUGHTON, R. The chemistry and applications of antimicrobial polymers: a state of-the-art review. *Biomacromolecules*. 2007, 8(5), pp. 1359-84. doi: 10.1021/bm061150q.
- 17.MAITZ, M. F. Applications of synthetic polymers in clinical medicine. *Biosurface and Biotribology*. 2015, **1**(3), pp. 161-176. doi:10.1016/j.bsbt.2015.08.002.
- 18.WALSH, G. *Proteins: Biochemistry and biotechnology*, 2nd ed. Oxford, UK: Willey-Blackwell, ©2014, ISBN: 978-0-470-66985-3.
- 19.SHAHZAD, M., MILLHOUSE, E., CULSHAW, S. Selected dietary (poly)phenols inhibit periodontal pathogen growth and biofilm formation. *Food & Function* 2015, **6**, pp. 719-729. doi: 10.1039/C4FO01087F.
- 20.YU, K., MEI, Y., HADJESFANDIARI, N. Engineering biomaterials surfaces to modulate the host response. *Colloids and Surfaces B: Biointerfaces*. 2014, **124**, pp. 69-79.
- 21.SIEDENBIEDEL, F., TILLER, J. C. Antimicrobial polymers in solution and on surfaces: Overview and functional principles. *Polymers*. 2012, **4**, pp. 46-71. doi:10.3390/polym4010046.
- 22.HUANG, K., YANG, Ch., HUANG, A. et al. Recent advances in antimicrobial polymers: A mini-review. *International Journal of Molecular Sciences*. 2016, **17**(9), 1578. doi: doi:10.3390/ijms17091578.
- 23.SHAHKARAMIPOUR, N., TRAN, T. N., RAMANAN, S., LIN, H. Membranes with surface-enhanced antifouling properties for water purification. *Membranes*. 2017, **7**(1), pp. 13. doi: 10.3390/membranes7010013.
- 24.BONILLA, J., ATARÉS, L., VARGAS, M., CHIRALT, A. Effect of essential oils and homogenization conditions on properties of chitosan-based films. Food Hydrocolloids. 2012, 26(1), pp. 9-16. doi: 10.1016/j.foodhyd.2011.03.015.
- 25. Size Theory [online]. © [viewed 2020-07-15]. Available from: https://www.chem.uci.edu/~dmitryf/manuals/Fundamentals/DLS%20mea surement%20principles.pdf.
- 26.SABBAH, M. T., Di PIERRO, P., ESPOSITO, M. et al. Stabilization of charged polysaccharide film forming solution by sodium chloride: Nanoparticle Z-average and zeta-potential monitoring. *Journal of*

- Biotechnology & Biomaterials. 2016, **6**(4), e128. doi: 10.4172/2155-952X.1000e128.
- 27.STAMM, M. *Polymer surfaces and interfaces: Characterization, modification and applications*. 1st ed. Berlin, Heidelberg: Springer-Verlag, ©2008. ISBN: 978-3-540-73864-0.
- 28.ROŠIC, R., PELIPENKO, J., KOCBEK, P. et al. The role of rheology of polymer solutions in predicting nanofiber formation by electrospinning. *European Polymer Journal*. 2012, **48**(8), pp. 1374-84. doi: 10.1016/j.eurpolymj.2012.05.001.
- 29.KRONBERG, B., HOLMBERG, K., LINDMAN, B. Surface chemistry of surfactants and polymers. West Sussex, UK: John Willey & Sons, Ltd. ©2014. ISBN: 9781119961246.
- 30.LIN, C.S., HORNG, T., CHEN, J. et al. Mechanical properties measurement of polymer films by bulge test and fringe projection. *Advances in Materials Science and Engineering*. 2014, 6. doi: 10.1155/2014/170279.
- 31.SEDLARIKOVA, J., JANALIKOVA, M., RUDOLF, O. et al. Chitosan/thyme oil systems as affected by stabilizing agent: physical and antimicrobial properties. *Coatings* 2019, **9**, pp. 165.
- 32.ALTIOK, D., ALTIOK, E., TIHMINLIOGLU, F. Physical, antibacterial and antioxidant properties of chitosan films incorporated with thyme oil for potential wound healing applications. *Journal of Materials Science: Materials in Medicine*. 2010, **21**(7), pp. 2227-36. doi: 10.1007/s10856-010-4065-x.
- 33.OJAGH, S. M., REZAEJ, M., RAZAVI, S. H. Development and evaluation of a novel biodegradable film made from chitosan and cinnamon essential oil with low affinity toward water. *Food Chemistry*. 2010, **122**(1), pp. 161-166. doi: 10.1016/j.foodchem.2010.02.033.
- 34.GOODWIN, J. W. Colloids and interfaces with surfactants and polymers an introduction. West Sussex, England: John Willey & Sons. ©2004. ISBN: 9780470093917.
- 35.GARCÍA, J. L., CUPESSALA, F., HUMPOLÍČEK, P., LEHOCKÝ, M. Physical and morphological changes of poly(tetrafluoroethylene) after using non-thermal plasma treatments. *Materials*. 2018, **11**(10), pp. 2013. doi: 10.3390/ma11102013.
- 36.KUN, E., MAROSSY, K. Evaluation methods of antimicrobial activity of plastics. *Materials Science Forum*. 2013, **729**, pp. 430-435. doi: 10.4028/www.scientific.net/MSF.729.430.
- 37.HUANG, S., RAS, R. H. A., TIAN, X. Antifouling membranes for oily wastewater treatment: Interplay between wetting and membrane fouling. *Current Opinion in Colloid and Interface Science*. 2018, **36**, pp. 90-109. doi: 10.1016/j.cocis.2018.02.002.

- 38.PEER, P., SEDLARIKOVA, J., JANALIKOVA, M., KUCEROVA, L., PLEVA, P. Novel poly vinylbutyral/monoacylglycerol nanofibrous membrane with antifouling activity. Accepted to *Materials*. 2020, **13**(7), 3662. doi: 10.3390/ma13173662.
- 39.GUZMÁN, E., LLAMAS, S., MAESTRO, A. Polymer–surfactant systems in bulk and at fluid interfaces. *Advances in Colloid and Interface Science*. 2016, **233**, pp. 38-64. doi: 10.1016/j.cis.2015.11.001.
- 40.REHMAN, N., KHAN, A., BIBI, I. Intermolecular interactions of polymer/surfactants mixture in aqueous solution investigated by various techniques. *Journal of Dispersion Science and Technology*. 2013, **34**(9), pp. 1202-1210. doi: 10.1080/01932691.2012.739940.
- 41.TAYLOR, D. J.F., THOMAS, R. K., PENFOLD, J. Polymer/surfactant interactions at the air/water interface. *Advances in Colloid and Interface Science* [online]. 2007, **132**(2), pp. 69-110. doi: 10.1016/j.cis.2007.01.002.
- 42.ULLAH, H., Ali, S. Classification of anti-bacterial agents and their functions. In: KUMAVATH, R. ed. *Antibacterial Agents*.. BoD-Books on Demand, 2017. ISBN 9535131990.
- 43.YANG, J., LEE, S., HAN, Y. Efficient transdermal penetration and improved stability of L-ascorbic acid encapsulated in an inorganic nanocapsule. *Bulletin-Korean Chemical Society*. 2003, **24**(4), pp. 499-503. doi: 10.5012/bkcs.2003.24.4.499.
- 44.HAMADOU, A. H., HUANG, W., XUE, Ch., MAO, X. Formulation of vitamin C encapsulation in marine phospholipids nanoliposomes: Characterization and stability evaluation during long term storage. *LWT*. 2020, **127**, pp. 109439. doi: 10.1016/j.lwt.2020.109439.
- 45.KHANNIRI, E., BAGHERIPOOR-FALLAH, SOHRABVANDI, S. et al. Application of liposomes in some dairy products. *Critical Rewievs in Food Science and Nutrition*. 2015, **56**(3), pp. 484-93. doi: 10.1080/10408398.2013.779571.
- 46.TEIXEIRA, M. C., SEVERINO, P., ANDREANI, T. et al. D-α-tocopherol nanoemulsions: Size properties, rheological behaviour, surface tension, osmolarity and cytotoxicity. *Saudi Pharmaceutical Journal*. 2017, **25**(2), pp. 231-235.
- 47. SCHRÖDER, A., LAGUERRE, M., SPRAKEL, J. et al. Pickering particles as interfacial reservoirs of antioxidants. *Journal of Colloid and Interface Science*. 2020, **575**, pp. 489-498. doi: 10.1016/j.jcis.2020.04.069.
- 48.GOËLO, V., CHAUMUN, M., GONCALVES, A. Polysaccharide-based delivery systems for curcumin and turmeric powder encapsulation using a spray-drying process. *Powder Technology*. 2020, **370**, pp. 137-146. doi: 10.1016/j.powtec.2020.05.016.
- 49.BASTOS, F., SANTOS, L. Encapsulation of cosmetic active ingredients for topical application a review. *Journal of Microencapsulation*. 2015, **33**(1), pp. 1-17. doi: 10.3109/02652048.2015.1115900.

- 50.LUXSUWONG, D., INDRANUPAKORN, R., WONGTRAKUL, P. Preparation of vesicles entrapped lycopene extract. *Journal of Oleo Science*. 2014, **63**(6), pp. 645-652. doi: 10.5650/jos.ess13216.
- 51.FRANCOLINI, I., DONELLI, G., STOODLEY, P. Polymer designs to control biofilm growth on medical devices. *Reviews in Environmental Science and Biotechnology*. 2003, **2**, 307-319.
- 52.MAHIRA, S., JAIN, A., KHAN, W., DOMB, A. J. Antimicrobial materials An overview. In *Antimicrobial Materials for Biomedical Applications*. 2019, pp. 1-37. doi: 10.1039/9781788012638-00001.
- 53. TAMPAU, A., GONZÁLEZ-MARTÍNEZ, Ch., CHIRALT, A. Polyvinyl alcohol-based materials encapsulating carvacrol obtained by solvent casting and electrospinning. *Reactive and Functional Polymers*. 2020, **153**, pp. 104603. doi: 10.1016/j.reactfunctpolym.2020.104603.
- 54.MAITZ, M. F. Applications of synthetic polymers in clinical medicine. *Biosurface and Biotribology*. 2015, **1**(3), pp. 161-176. doi: 10.1016/j.bsbt.2015.08.002.
- 55.OLEWNIK-KRUSZKOWSKA, E., GIERSZEWSKA, M., JAKUBOWSKA, E. et al. Antibacterial films based on PVA and PVA-chitosan modified with poly(hexamethylene guanidine). *Polymers*. 2019, **11**(12), pp. 2093. 10.3390/polym11122093.
- 56.HU, D., WANG, L. Preparation and characterization of antibacterial films based on polyvinyl alcohol/quaternized cellulose. *Reactive and Functional Polymers*. 2016, **101**, pp. 90-98. doi: 10.1016/j.reactfunctpolym.2016.02.012.
- 57.SHINTANI, H. Modification of medical device surface to attain Anti-Infection. *Trends in Biomaterials and Artificial Organs*. 2004, **18**(1), pp. 1-8.
- 58.LIANG, X., WANG, A., CAO, T. effect of cast molded rifampicin/silicone on Staphylococcus epidermidis biofilm formation. *Journal of Biomedical Materials Research Part A.* 2006, **76**(3), pp. 580-8. doi: 10.1002/jbm.a.30559.
- 59.MERCHAN, M., SEDLARIKOVA, J., SEDLARIK, V., MACHOVSKY, M., SVOBODOVA, J., SAHA, P. Antibacterial polyvinyl chloride/antibiotic films: The effect of solvent on morphology, antibacterial activity and release kinetics. *Journal of Applied Polymer Science* 118, 2010, 2369-2378.
- 60.SIEMANN, U. Solvent cast technology a versatile tool for thin film production. Berlin, Heidelberg: Springer-Verlag ©2005.In: Stribeck N., Smarsly B. (eds) Scattering Methods and the Properties of Polymer Materials. Progress in Colloid and Polymer Science. ISBN 978-3-540-25323-5.

- 61.DUTA, O. C., MAXIMOV, M., TRUSCA, R. et al. Advanced drug-eluting poly(vinyl chloride) surfaces deposited by spin coating. *Medicina*. 2019, **55**(8), pp. 421. doi: 10.3390/medicina55080421.
- 62.KUMAR, R., MÜNDSTEDT, H. Silver ion release from antimicrobial polyamide/silver composites. *Biomaterials*. 2005, **26**(14), pp. 2081-2088. doi: 10.1016/j.biomaterials.2004.05.030.
- 63.Ampicillin Sodium Salt. *Sigma Aldrich* [online]. © [viewed 2020-07-20]. Available from: https://www.sigmaaldrich.com/catalog/product/sigma/a8351?lang=en®ion=CZ.
- 64.YEDDES, W., DJEBALI, K., WANNES, W. A. et al. Gelatin-chitosan-pectin films incorporated with rosemary essential oil: optimized formulation using mixture design and response surface methodology. *International Journal of Biological Macromolecules*. 2020, **154**, 92-103. doi: 10.1016/j.ijbiomac.2020.03.092.
- 65.DU, W., AVENA-BUSTILLOS, R. J., Hua, S. S. T., McHugh, T. H. Antimicrobial volatile essential oils in edible films for food safety. *Science against Microbial Pathogens: Communicating Current Research and Technological Advances*. 2011, pp. 1124–1134.
- 66.SIRIPATRAWAN, U., NOIPHA, S. Active film from chitosan incorporating green tea extract for shelf life extension of pork sausages. *Food Hydrocolloids*. 2012, **27**(1), pp. 102–108. doi: 10.1016/j.foodhyd.2011.08.011.
- 67.PRANOTO, Y., RAKSHIT, S. K., SALOKHE, V. M. Enhancing antimicrobial activity of chitosan films by incorporating garlic oil, potassium sorbate and nisin. *LWT-Food Science and Technology*. 2005, **38**(8), pp. 859–865, 2005. doi: 10.1016/j.lwt.2004.09.014.
- 68.WANG, L., LIU, F. Jiang, Y. et al. Synergistic antimicrobial activities of natural essential oils with chitosan films. *Journal of Agricultural and Food Chemistry*. 2011, **59**(23), pp. 12411–12419.
- 69.ABDOLLAHI, M., Rezaei, M., Farzi, G. Improvement of active chitosan film properties with rosemary essential oil for food packaging. *International Journal of Food Science and Technology*. 2012, **47**(4), pp. 847–853. doi: 10.1111/j.1365-2621.2011.02917.x.
- 70.OZCALIK, O., TIHMINLIOGLU, F. Barrier properties of corn zein nanocomposite coated polypropylene films for food packaging applications. *Journal of Food Engineering*. 2013, **114**(), pp. 505-513. doi:10.1016/J.JFOODENG.2012.09.005.
- 71.KASHIRI, M., CERISUELO, J. P., DOMÍNGUEZ, I. B. et al. Novel antimicrobial zein film for controlled release of lauroyl arginate (LAE). *Food Hydrocolloids*. 2016, **61**(), pp. 547-554. doi: 10.1016/J.FOODHYD.2016.06.012.

- 72.Del NOBILE, M. A., CONTE, A., INCORONATO, A. L., PANZA, O. Antimicrobial efficacy and release kinetics of thymol from zein films. *Journal of Food Engineering*. 2008, **89**(1), pp. 57-63. doi: 10.1016/j.jfoodeng.2008.04.004.
- 73.MASTROMATTEO, M., BARBUZZI, G., Del NOBILE, M. A., Controlled release of thymol from zein based films. *Innovative Food Science & Emerging Technologies*. 2009, **10**(2), pp. 222-227. doi: 10.1016/j.ifset.2008.11.010.
- 74.SINGH, N., GEORGET, D. M. R., BELTON, P., BARKER, S. Physical properties of zein films containing salicylic acid and acetyl salicylic acid. *Journal of Cereal Science*. 2010, **52**(2), p. 282-287. doi: 10.1016/j.jcs.2010.06.008.
- 75.JANES, M. E., KOOSHESH, S., JOHNSON, M. G. Control of Listeria monocytogenes on the surface of refrigerated, ready-to-eat chicken coated with edible zein film coatings containing nisin and/or calcium propionate. *Journal of Food Science*. 2006, 67(7), pp. 2754-2757. doi: 10.1111/j.1750-3841.2007.00633.x.
- 76.PETLIN, D., TVERDOKHLEBOV, S. I., ANISSIMOV, Y. G. Plasma treatment as an efficient tool for controlled drug release from polymeric materials: A review. *Journal of Controlled Release*. 2017, **266**, pp. 57-74, doi: 10.1016/j.jconrel.2017.09.023.
- 77.PARADISO, P., CHU, V. et al. Effect of plasma treatment on the performance of two drug-loaded hydrogel formulations for therapeutic contact lenses. *Journal of Biomedical Materials Research Part B Applied Biomaterials*. 2015, **103**(5), pp. 1059-1068. doi: 10.1002/jbm.b.33287.
- 78.McINNES, S. J. P., MICHI, T. D., DELALAT, B. et al. "Thunderstruck": plasma-polymer-coated porous silicon microparticles as a controlled drug delivery system. *ACS Applied Materials and Interfaces*. 2016, **8** (7), pp. 4467-4476. doi: 10.1021/acsami.5b12433.
- 79.YAMAUCHI, Y. Y. Yamauchi, et al. Construction of matrix-type drug delivery system using solid phase polymerization initiated by plasma-induced radicals. *Journal of Photopolymer Science and Technology*, 2013 **26** (4), pp. 529-532. doi: 10.2494/photopolymer.26.529.
- 80.SARAF, A., JOHNSON, K., LIND, M.L. Poly(vinyl) alcohol coating of the support layer of reverse osmosis membranes to enhance performance in forward osmosis. *Desalination*. 2014, **333**, pp. 1-9. doi: 10.1016/j.desal.2013.11.024.
- 81.FRIEDRICH, J. *The plasma chemistry of polymer surfaces: Advanced techniques for surface design*. Weinheim, Germany: Wiley-VCH Verlag & Co. KGaA (1st ed). ©2012. ISBN: 978-3-527-31853-7.
- 82.CHOUKOUROV, A., HANUŠ, J., KOUSAL, J. et al. Thin polymer films from polyimide vacuum thermal degradation with and without a glow

- discharge. *Vacuum*. 2006, **80**(8), pp. 923-929. doi: 10.1016/j.vacuum.2005.12.012.
- 83.PEER, P., STENICKA, M., PAVLINEK, V. An electrorheological investigation of PVB solutions in connection with their electrospinning qualities. *Polymer Testing*. 2014, **39**, pp. 11-121. doi: 10.1016/j.polymertesting.2014.07.016.
- 84.KURTZ, I. S., SCHIFFMAN, J. D. Current and emerging approaches to engineer antibacterial and antifouling electrospun nanofibers. *Materials*. 2018, **11**(7), pp. 1059, doi: doi: 10.3390/ma11071059.
- 85.CAI, N., LI, C., HAN, C. et al. Tailoring mechanical and antibacterial properties of chitosan/gelatinnanofiber membranes with Fe3O4 nanoparticles for potential wound dressing application. *Applied Surface Science*. 2016, **369**, pp. 492-500. doi: 10.1016/j.apsusc.2016.02.053.
- 86.PEER, P., POLASKOVA, M., MUSILVA, L. Superhydrophobic poly(vinyl butyral) nanofibrous membrane containing various silica nanoparticles. *Journal of the Textile Institute*. 2019, **110**, pp. 1508-1514. doi: 10.1080/00405000.2019.1605658.
- 87.GE, L., ZHAO, Y.S., MO, T. et al. Immobilization of glucose oxidase in electrospun nanofibrous membranes for food preservation. *Food Control*. 2012, **26**, pp. 88–193.
- 88.RASOULI, R., BARHOUM, A., BECHELANY, M.; DUFRESNE, A. Nanofibers for biomedical and healthcare applications. *Macromolecular Bioscience*. 2019, **19**, pp. 1800256. doi: 10.1002/mabi.201800256.
- 89.RIVERO, P.J., URRUTIA, A., GOICOECHEA, J. et al. Nanomaterials for functional textiles and fibers. *Nanoscale Research Letters*. 2015, **10**, pp. 501. doi: 10.1186/s11671-015-1195-6.
- 90.SHAHKARAMIPOUR, N., TRAN, T.N., RAMANAN, S. et al. Membranes with surface-enhanced antifouling properties for water purification. *Membranes* 2017, **7**(1), pp. 13. doi: 10.3390/membranes7010013.
- 91.HU, M., LI, C.W., Li, X. et al. Zinc oxide/silver bimetallic nanoencapsulated in PVP/PCL nanofibers for improved antibacterial activity. *Artificial Cells Nanomedicine and Biotechnology*. 2018, **46**, pp. 1248-1257. doi: 10.1080/21691401.2017.1366339.
- 92.RIEGER, K.A., SCHIFFMAN, J.D. Electrospinning an essential oil: Cinnamaldehyde enhances the antimicrobial efficacy of chitosan/poly(ethylene oxide) nanofibers. *Carbohydrate Polymers*. 2014, **113**, pp. 561-568. doi: 10.1016/j.carbpol.2014.06.075.
- 93.FAN, X.Y., YIN, M.L., JIANG, Z.M. et al. Antibacterial poly(3-hydroxybutyrate-co-4-hydroxybutyrate) fibrous membranes containing quaternary ammonium salts. *Polymers for Advanced Technologies*. 2016, **27**, pp. 1617-1624. doi: 10.1002/pat.3839.

- 94.PARK, J.A., KIM, S.B. Anti-biofouling enhancement of a polycarbonate membrane with functionalized poly(vinyl alcohol) electrospun nanofibers: Permeation flux, biofilm formation, contact, and regeneration tests. *Journal of Membrane Science*. 2017, **540**, pp. 192–199. doi: 10.1016/j.memsci.2017.06.071.
- 95.FRANCOLINI, I., VUOTTO, C., PIOZZI, A., DONELLI, G. Antifouling and antimicrobial biomaterials: an overview. *APMIS*. 2017, **125**, pp. 392-417. doi: 10.1111/apm.12675.
- 96.PEER, P., STENICKA, M., PAVLINEK, V., FILIP, P. The storage stability of polyvinylbutyral solutions from an electrospinnability standpoint. *Polymer Degradation and Stability*. 2014, **10**(1), pp. 134-139, doi: 10.1016/j.polymdegradstab.2014.04.015.
- 97.KABARA, J.J. Lipids as host-resistance factors of human milk. *Nutrition Reviews*. 1980, **38**(2), pp. 65-73. doi: 10.1111/j.1753-4887.1980.tb05843.x.
- 98.THORMAR, H., HILMARSSON, H. Glycerol monocaprate (monocaprin) reduces contamination by Escherichia coli and Salmonella enteritidis in hard surfaces. *Food Control*. 2012, **25**, pp. 505-510. doi: 10.1016/j.foodcont.2011.11.024.
- 99.ARAUJO, E.S.; NASCIMENTO, M.L.F.; de OLIVIERA, H.P. Influence of triton X-100 on PVA fibers production by the electrospinning technique. *Fibres and Textiles in Eastern Europe*. 2013, **100**, pp. 39-43.

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LIST OF SYMBOLS AND ABBREVIATIONS

AgCl silver chloride AgNO₃ silver nitrate

ATR-FTIR attenuated total reflectance-Fourier transform infrared spectroscopy

BHA butylated hydroxyanisole BHT butylated hydroxytoluene

CAC critical aggregation concentration

CFU colony forming units

CMC critical micellar concentration

CYH cyclohexanone

DCSBD dielectric coplanar surface barrier discharge

DLS dynamic light scattering
DMF N, N-dimethylformamide
DNA deoxyribonucleic acid
DSS dioctyl sulfosuccinate
EIP emulsion inversion point

EO essential oil

GUV giant unilamellar vesicles HLB hydrophilic-lipophilic balance

HPLC-UV high performance liquid chromatography with UV detection

LUV large unilamellar vesicles

MAG C10/C12 monoacylglycerol of capric/lauric acid MIC minimum inhibition concentration

MLV multilamellar vesicles

MRSA methicillin resistant *Staphylococcus aureus*

NLC nanostructured lipid carriers

OLV oligolamellar vesicles O/W oil in water emulsion

PAVTD plasma assisted vacuum thermal deposition

PCS photon correlation spectroscopy

PECVD plasma-enhanced chemical vapour deposition

PEO poly(ethylene oxide)

PIT phase inversion temperature

PPO poly(propylene oxide)
PVA poly(vinyl alcohol)
PVB poly(vinyl butyral)
PVC poly(vinyl chloride)

SEM scanning electron microscopy
SLN solid lipid nanoparticles
SUV small unilamellar vesicles

TEM transmission electron microscpy

W/O water in oil emulsion

WVP water vapour permeability

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1. JANIS, R, KLASEK, A, KREJCI, J, BOBALOVA, J. Influence of some chromium complexes on the conversion rate of glycidol - Fatty acid reaction. *Tenside Surfactants Detergents* 2005, 42(1), 44-48. DOI: 10.3139/113.100250.

2. JANIS, R, KLASEK, A, BOBALOVA, J. Chromium (III) acetate hydroxide - An efficient catalyst for preparation of 1-monoacylglycerols by

- the glycidol-fatty acid reaction. *Journal of Food Lipids* 2006, 13(2), 199-209. DOI: 10.1111/j.1745-4522.2006.00045.x.
- 3. GALYA, T, SEDLARIK, V, KURITKA, I, SEDLARIKOVA, J, SAHA, P. Characterization of antibacterial polymeric films based on poly(vinyl alcohol) and zinc nitrate for biomedical applications. *International Journal of Polymers Analysis and Characterization* 2008, 13(4), 241-253. DOI: 10.1080/10236660802175790.
- 4. GALYA, T, SEDLARIK, V, KURITKA, I, NOVOTNY, R, SEDLARIKOVA, J, SAHA, P. Antibacterial Poly(vinyl Alcohol) Film Containing Silver Nanoparticles: Preparation and Characterization: Preparation and Characterization. *Journal of Applied Polymer Science* 2008, 110(5), 3178-3185. DOI: 10.1002/app.28908.
- 5. SEDLARIK, V, GALYA, T, SEDLARIKOVA, J, VALASEK, P, SAHA, P. The Effect of Hydrolysis Degree on the Properties of Antibacterial Polymeric Films Based on Poly(vinyl alcohol) and Zinc Sulphate for Biomedical Applications. *Journal of Biomaterials Science-Polymer Edition* 2010, 21(11), 1421-1440. DOI: 10.1163/092050609X12517858243742.
- 6. MERCHAN, M, SEDLARIKOVA, J, VESEL, A, SEDLARIK, V, PASTOREK, M, SAHA, P. Characterization of Antibacterial, Mechanical, and Structural Properties of Polyvinyl Chloride/Silver Nitrate Composites Prepared by Thermoplastic Compounding. *International journal of Polymer Analysis and Characterization* 2010, 15(6), 360-369. DOI: 10.1080/1023666X.2010.500534.
- 7. SEDLARIK, V, GALYA, T, SEDLARIKOVA, J,VALASEK, P, SAHA, P. The effect of preparation temperature on the mechanical and antibacterial properties of poly(vinyl alcohol)/silver nitrate films. *Polymer Degradation and Stability* 2010, 95(3), 399-404. DOI: 10.1016/j.polymdegradstab.2009.11.017.
- 8. MERCHAN, M, SEDLARIKOVA, J, SEDLARIK, V, MACHOVSKY, M, SVOBODOVA, J, SAHA, P. Antibacterial Polyvinyl Chloride/Antibiotic Films: The Effect of Solvent on Morphology, Antibacterial Activity, and Release Kinetics: The Effect of Solvent on Morphology, Antibacterial Activity, and Release Kinetics. *Journal of Applied Polymer Science* 2010, 118(4), 2369-2378. DOI: 10.1002/app.32185.
- 9. MERCHAN, M, SEDLARIKOVA, J, FRIEDRICH, M, SEDLARIK, V, SAHA, P. Thermoplastic modification of medical grade polyvinyl chloride with various antibiotics: effect of antibiotic chemical structure on mechanical, antibacterial properties, and release activity: effect of antibiotic chemical structure on mechanical, antibacterial properties, and release activity. *Polymer Bulletin* 2011, 67(6), 997-1016. DOI: 10.1007/s00289-011-0474-3.

- 10. MERCHAN, M, SEDLARIKOVA, J, VESEL, A, MACHOVSKY, M, SEDLARIK, V, SAHA, P. Antimicrobial Silver Nitrate-doped Polyvinyl Chloride Cast Films: Influence of Solvent on Morphology and Mechanical Properties: Influence of Solvent on Morphology and Mechanical Properties. *International Journal of Polymeric Materials and Polymeric Biomaterials* 2013, 62(2), 101-108. DOI: 10.1080/00914037.2012.670821.
- 11. EGNER, P, KASPARKOVA, V, PAVLACKOVA, J, SEDLARIKOVA, J, PINDAKOVA, L. Effect of process parameters and methylcellulose supplementation on the properties of n-undecane emulsions. *Journal of Dispersion Science and Technology* 2017, 38(6), 775-781. DOI: 10.1080/01932691.2016.1194213.
- 12. SEDLARIKOVA, J, DOLEZALOVA, M, EGNER, P, PAVLACKOVA, J, KREJCI, J, RUDOLF, O, PEER, P. Effect of Oregano and Marjoram Essential Oils on the Physical and Antimicrobial Properties of Chitosan Based Systems. *International Journal of Polymer Science* 2017. DOI: 10.1155/2017/2593863.
- 13. PAVLACKOVA, J, KOVACSOVA, K, RADIMERSKY, P, EGNER, P, SEDLARIKOVA, J, MOKREJS, P. Stability and in vivo efficiency of natural cosmetic emulsion systems with the addition of vegetable oils. *Brazilian Journal of Pharmaceutical Sciences* 2018, 54(3). DOI: 10.1590/s2175-97902018000317693.
- 14. RASKOVA, ZK, STAHEL, P, SEDLARIKOVA, J, MUSILOVA, L, STUPAVSKA, M, LEHOCKY, M. The Effect of Plasma Pretreatment and Cross-Linking Degree on the Physical and Antimicrobial Properties of Nisin-Coated PVA Films. *Materials* 2018, 11(8). DOI: 10.3390/ma11081451.
- 15. PAVLACKOVA, J, EGNER, P, SEDLACEK, T, MOKREJS, P, SEDLARIKOVA, J, POLASKOVA, J. In vivo efficacy and properties of semisolid formulations containing panthenol. *Journal of Cosmetic Dermatology* 2019, 18(1), 346-354. DOI: 10.1111/jocd.12527.
- 16. SEDLARIKOVA, J, JANALIKOVA, M, RUDOLF, O, PAVLACKOVA, J, EGNER, P, PEER, P, V VARAD'OVA, V, KREJCI, J. Chitosan/Thyme Oil Systems as Affected by Stabilizing Agent: Physical and Antimicrobial Properties: Physical and Antimicrobial Properties. *Coating* 2019, 9(3). DOI: 10.3390/coatings9030165.
- 17. KOUSAL, J, KRTOUS, Z, RASKOVA, ZK, SEDLARIKOVA, J. et al. Degradable plasma polymer films with tailored hydrolysis behavior. *Vacuum* 2020, 173. DOI: 10.1016/j.vacuum.2019.109062.
- 18. KOUSAL, J, SEDLARIKOVA, J,KOLAROVA-RASKOVA, ZK et al. Degradable Poly(ethylene oxide)-Like Plasma Polymer Films Used for the Controlled Release of Nisin. *Polymers* 2020, 12(6). DOI: 10.3390/polym12061263.

19. PEER, P, SEDLARIKOVA, J, JANALIKOVA, M, KUCEROVA, L, PLEVA, P. Novel poly vinylbutyral/monoacylglycerol nanofibrous membrane with antifouling activity. *Materials* 2020, 13, 3662.

List of research projects:

- 1. GA17-10813S Novel plasma polymers with tunable stability and permeability, 2017–2019, Grant Agency of the Czech Republic, collaborator
- 2. OP VK CZ.1.07_2.2.00_28.0132 Increase of exclusiveness in education of fats, cosmetics and detergents technology (Zvyšování exkluzivity výuky technologie tuků, kosmetiky a detergentů). 2012-2015, collaborator
- 3. MEO9072 Study of polylactide based material for packaging applications. 2009-2012, Ministry of Education, Youth and Sports, collaborator
- 4. MEB090803 Research on thin polymeric layers for biomedical applications, 2008-2009, Ministry of Education, Youth and Sports, collaborator
- 5. 2A-1TP1/126 Innovation of Medical Polymeric Aids. 2006 2011, Ministry of Industry and Trade, collaborator
- 6. MSM7088352101 Multifunctional Composite Systems Based on Natural and synthetic Polymers. 2005-2010, Ministry of Education, Youth and Sports, collaborator
- 7. G1RD-CT-2002-00772 RTD Radial Environmentally Tannery Operation by Resource Management RESTORM. 2001-2006, collaborator

PUBLICATIONS

The full texts of publications P1 to P13 discussed within the text of habilitation thesis are provided in the following part.

Ing. Jana Sedlaříková, Ph.D.

Polymers as Carriers of Active Agents

Polymery jako nosiče aktivních látek

Habilitation Thesis

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